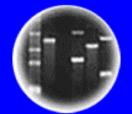




Entire Genetic Code of a Bacteria



DNA Fingerprinting



Cloning: Ethical Issues and Future Consequences



Plants of Tomorrow

HC70A & SAS70A Winter 2011 Genetic Engineering in Medicine, Agriculture, and Law

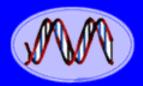
Professors Bob Goldberg & John Harada

Lecture 4

The Nuts & Bolts of Genetic Engineering: The Factor VIII Story -From Gene To Drug

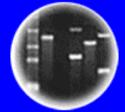








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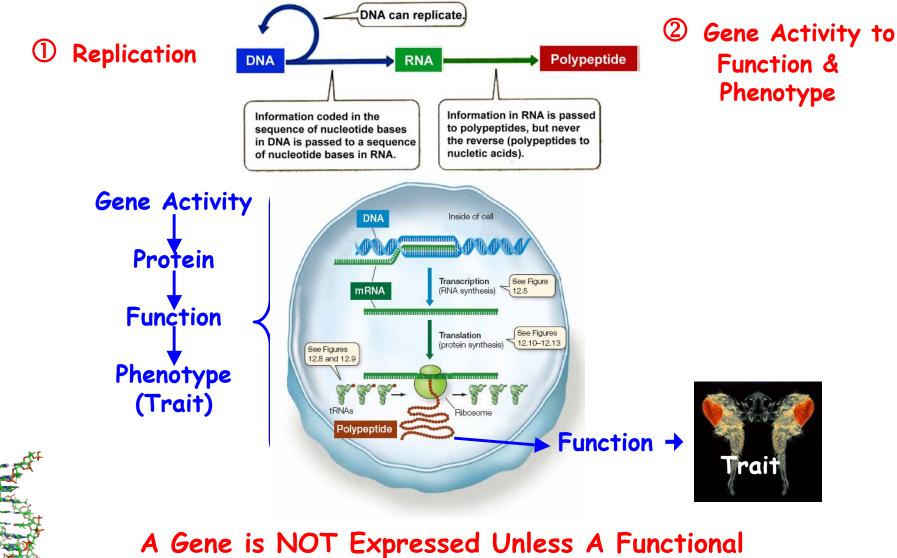


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THEMES

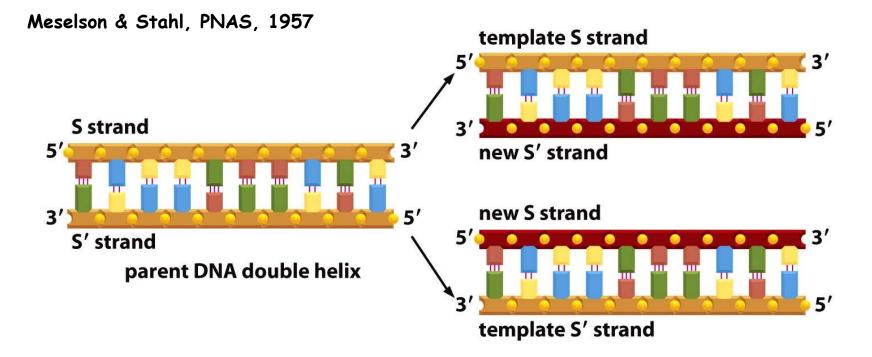
- 1. Continuation of How Do Genes Work Parts One and Two
- 2. What is Hemophilia?
- 3. How Is Hemophilia Inherited?
- 4. What is the Pedigree Pattern of a Sex-Linked Gene?
- 5. How Find a Disease Gene When It is Not Known Where the Gene is Expressed?
- 6. What Vectors Can Be Used For Cloning DNA?
- 7. What Are the Advantage of Using a Virus Vector For Constructing Genome Libraries?
- 8. How Make a Library of the Human Genome?
- 9. How Find a Gene With Only a Knowledge of the Protein Sequence?
- 10. What is Chromosome Walking & What Role Did it Play in Cloning the Factor VIII Gene?
- 11. How Use DNA Testing to Detect Factor VIII Disease Alleles?
- 12. How Isolate a Factor VIII cDNA Clone?
- 13. How Produce Factor VIII Protein For Use as a Drug?

How Do Genes Work (Lecture 3 Continued)



Protein Produced!

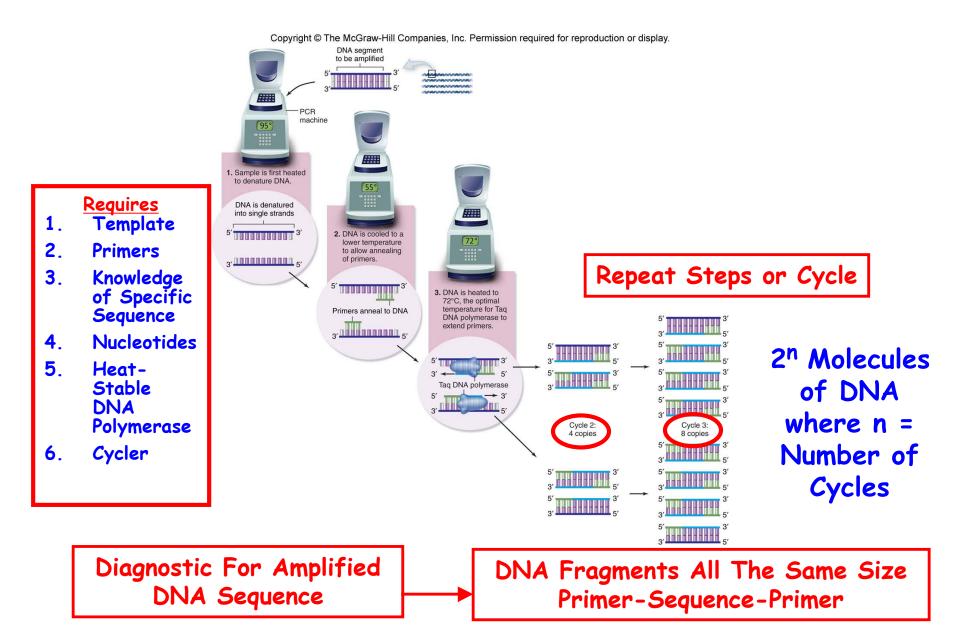
DNA Replication Occurs Semi-Conservatively



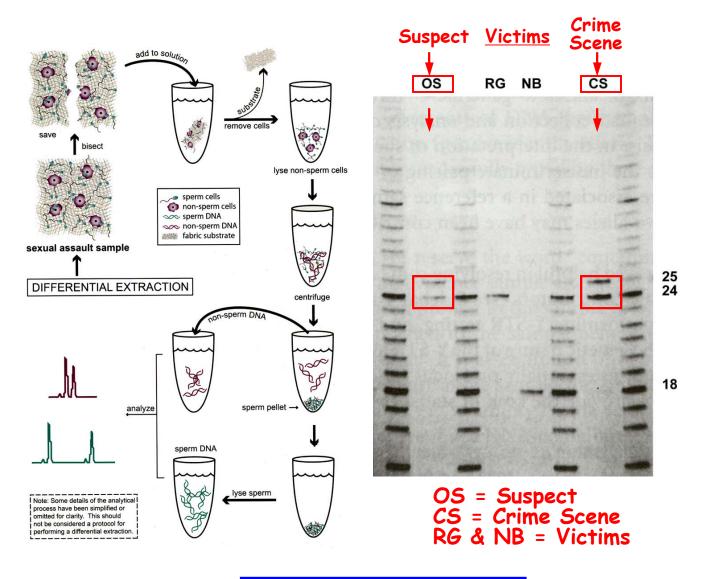
- 1. DNA Structure Allows DNA Sequence to Be Maintained by Complementary Base Pairing
- 2. Each Strand Serves as a Template for the Synthesis of a Complementary Strand
- New DNA Molecules are Precise Copies of Parental DNA

 Each Containing One Newly Synthesized Complementary
 Strand

PCR is A Cyclical Process of DNA Replication



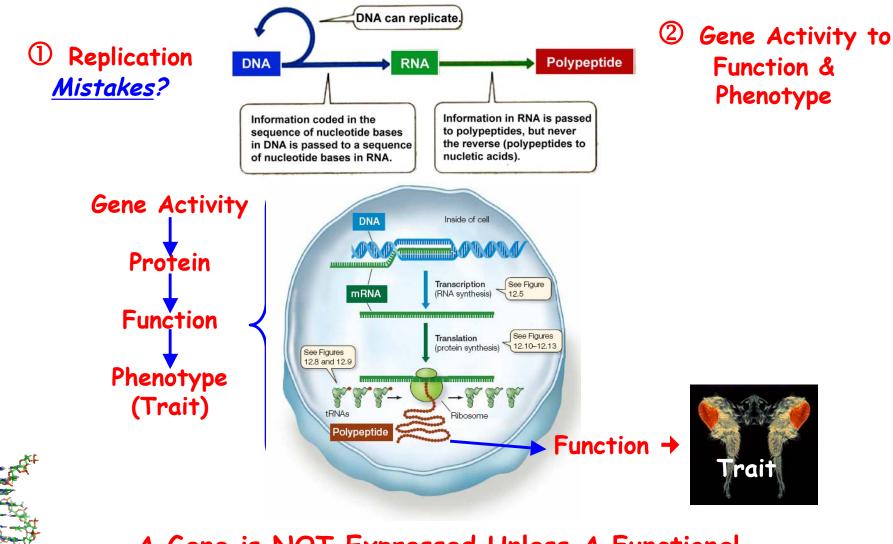
Using PCR in Crime Scenes



"Match" What is Probability That This Will Occur by Chance?

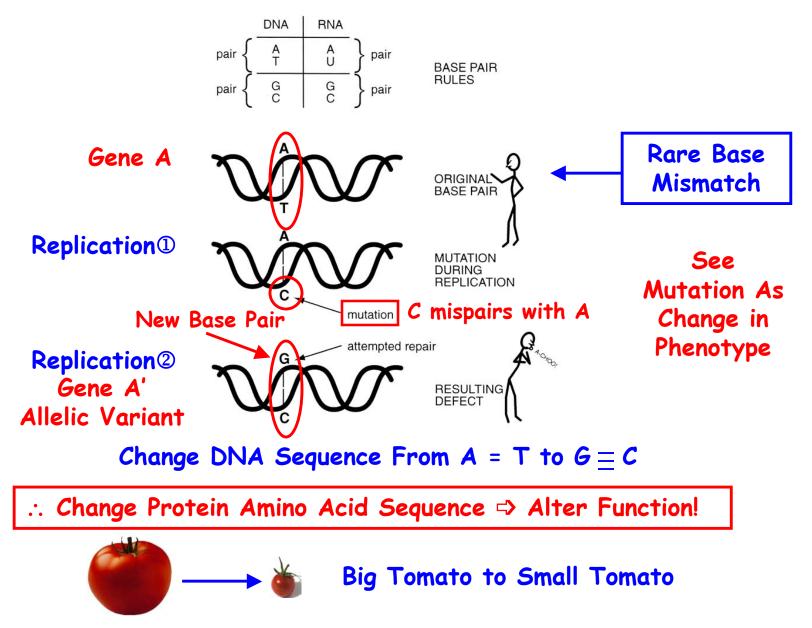
DNA Doesn't "Lie" !!

How Do Genes Work - Mutations!

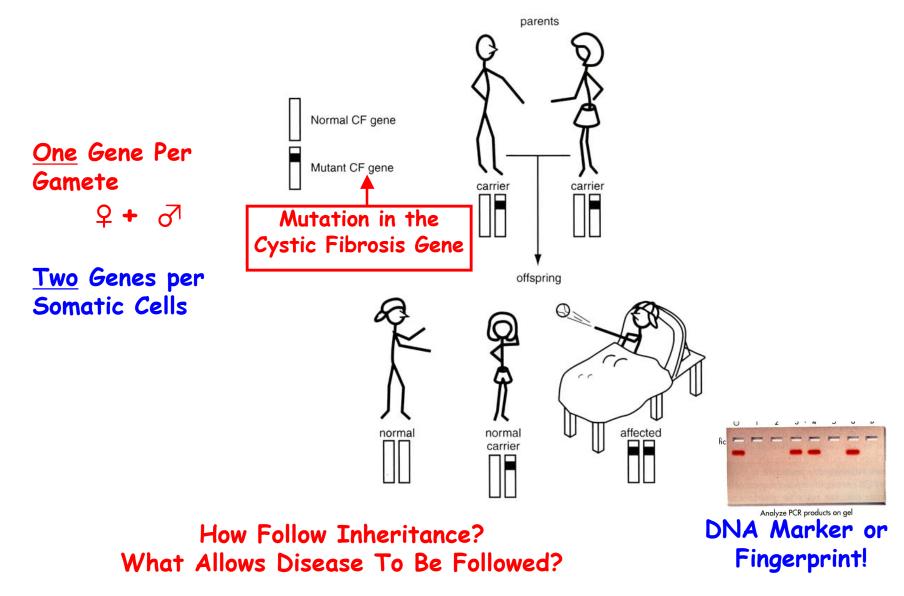


A Gene is NOT Expressed Unless A Functional Protein Produced!

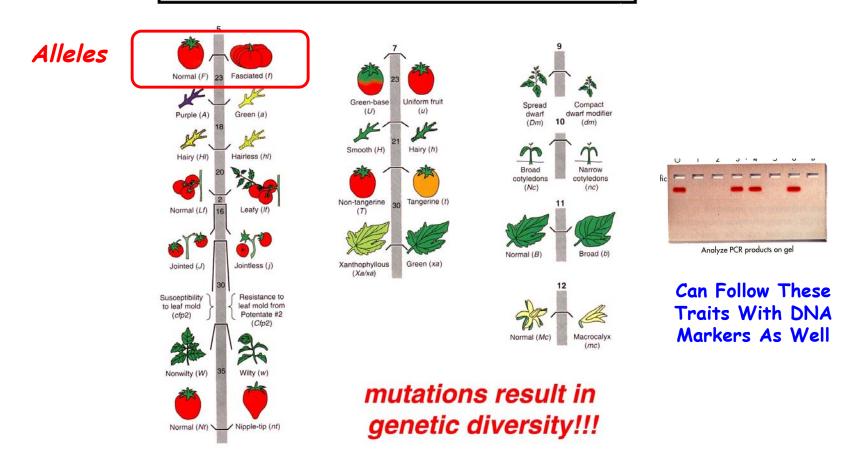
DNA Replication is Precise But Mistakes or Mutations Can Occur!!



Mutation in Genes Are Rare But Are Inherited (1 out of 10⁷ replications)

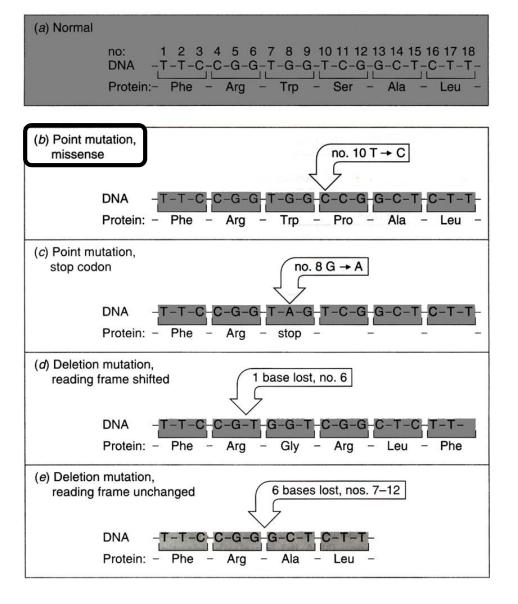


Alternative Forms of the Same Gene Lead to Genetic Diversity

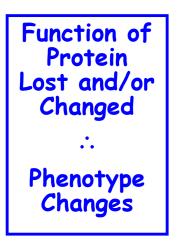


Spontaneous Mutations Give Rise To Alleles, or Different Forms of the Same Gene, And result in Small DNA Sequence Changes (e.g., SNPs or Single Nucleotide Polymorphisms)

Mutations Can Occur Different Ways

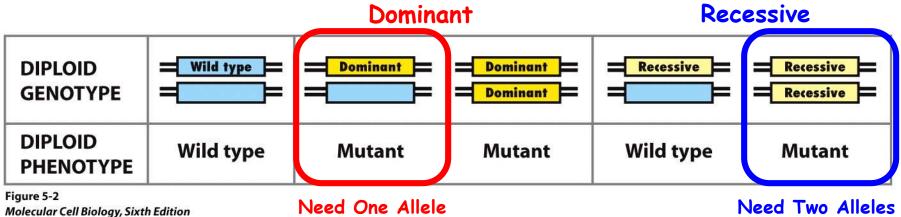


- 1. Base-Pair Change
- 2. Insert or Delete Base (Indel)
- 3. Move Gene, or Part of Gene, to New Location (Switches Change)!

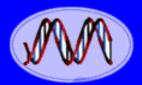


Human Genetic Disorders Occur As a Result of Mutations

TABLE 13.2	3.2 Some Important Genetic Disorders				
Disorder	Symptom	Defect	Dominant/ Recessive	Frequency Among Human Births	
Hemophilia	Blood fails to clot	Defective blood-clotting factor VIII	X-linked recessive	1/10,000 (Caucasian males)	
Huntington disease	Brain tissue gradually deteriorates in middle age	Production of an inhibitor of brain cell metabolism	Dominant	1/24,000	
Muscular dystrophy (Duchenne)	Muscles waste away	Degradation of myelin coating of nerves stimulating muscles	X-linked recessive	1/3700 (males)	
Hypercholesterolemia	Excessive cholesterol levels in blood lead to heart disease	Abnormal form of cholesterol cell surface receptor	Dominant	1/500	

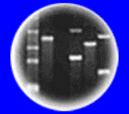


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Plants of Tomorrow

ARTICLE

Nature, October 10, 2010

doi:10.1038/nature09534

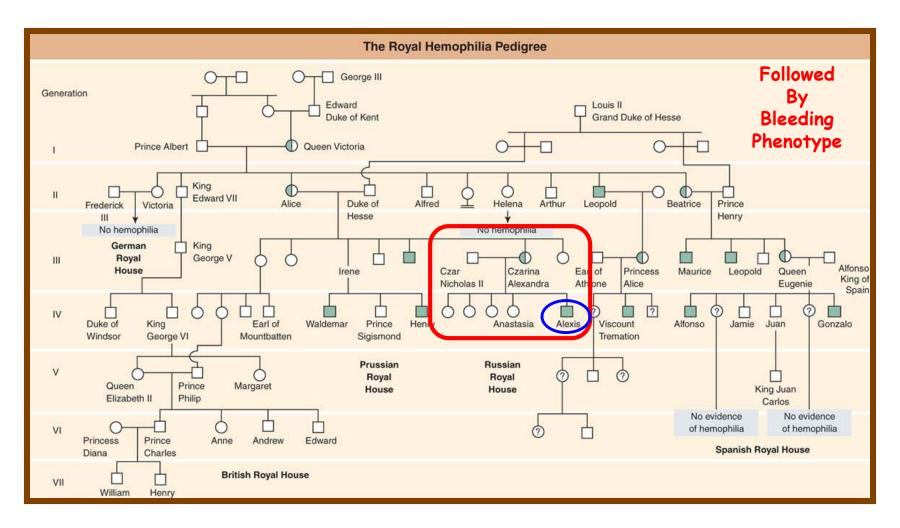
A map of human genome variation from population-scale sequencing

The 1000 Genomes Project Consortium*

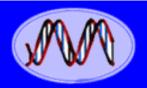
The 1000 Genomes Project aims to provide a deep characterization of human genome sequence variation as a foundation for investigating the relationship between genotype and phenotype. Here we present results of the pilot phase of the project, designed to develop and compare different strategies for genome-wide sequencing with high-throughput platforms. We undertook three projects: low-coverage whole-genome sequencing of 179 individuals from four populations; high-coverage sequencing of two mother-father-child trios; and exon-targeted sequencing of 697 individuals from seven populations. We describe the location, allele frequency and local haplotype structure of approximately 15 million single nucleotide polymorphisms, 1 million short insertions and deletions, and 20,000 structural variants, most of which were previously undescribed. We show that, because we have catalogued the vast majority of common variation, over 95% of the currently accessible variants found in any individual are present in this data set. On average, each person is found to carry approximately 250 to 300 loss-of-function variants in annotated genes and 50 to 100 variants previously implicated in inherited disorders. We demonstrate how these results can be used to inform association and functional studies. From the two trios, we directly estimate the rate of *de novo* germline base substitution mutations to be approximately 10^{-8} per base pair per generation. We explore the data with regard to signatures of natural selection, and identify a marked reduction of genetic variation in the neighbourhood of genes, due to selection at linked sites. These methods and public data will support the next phase of human genetic research.

- Sequenced Genomes of ~900 individuals
- From Seven Different Global Populations
- Found 250-300 Loss-Of-Function Mutations (KOs) Per Person
- 10⁻⁸ bp Mutations per Generation (30 per Genome)

Pedigrees Can Be Used To Follow Disease Genes in Human Families

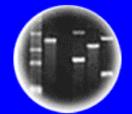


Recessive Sex Linked





Entire Genetic Code of a Bacteria



DNA Fingerprinting



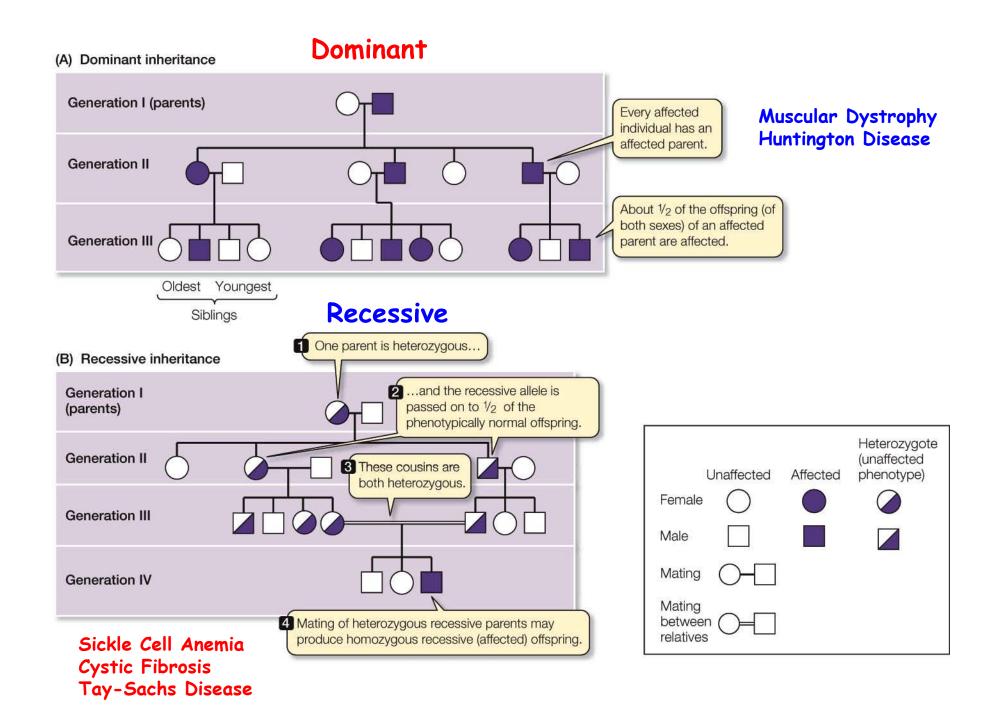
Cloning: Ethical Issues and Future Consequences



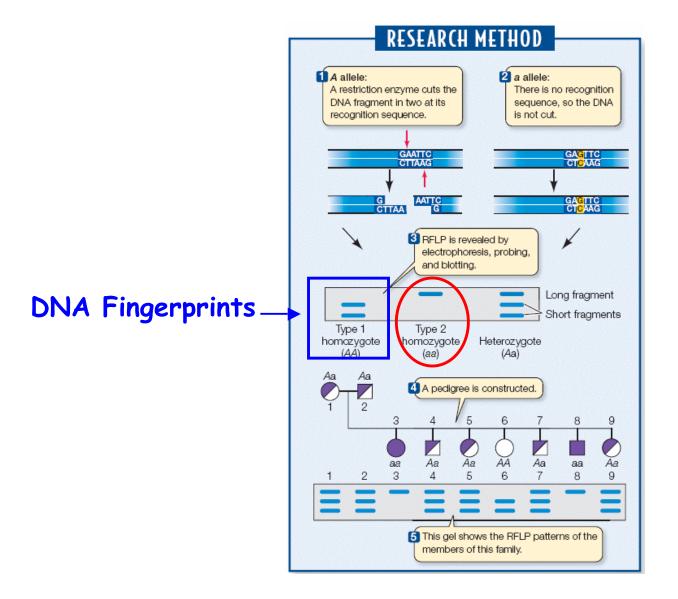
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Pedigrees Can Be Used To Determine If a Trait is Dominant or Recessive

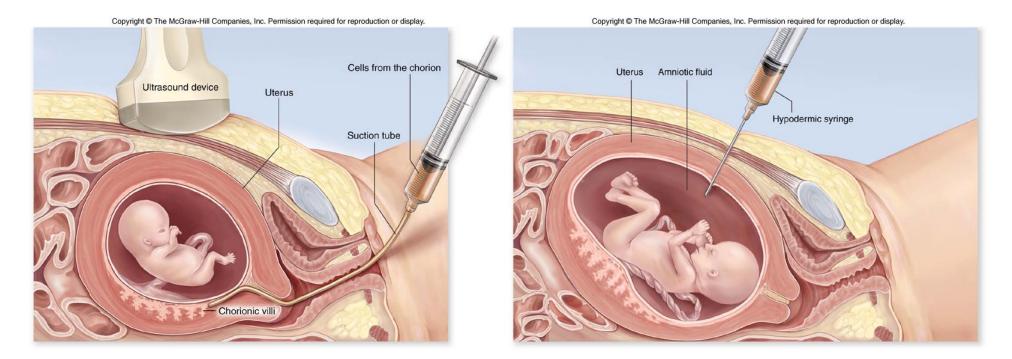
Each Type of Inheritance Predicts Specific Results in Each Generation



Genetic Diseases Can Be Followed in Families Using Molecular Methods (e.g., DNA Blots or PCR)

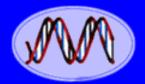


PCR Can Be Used To Analyze Genes During Pregnancy



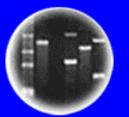
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TABLE 13.2	Some Important Genetic Disorders					
Disorder	Symptom	Defect	Dominant/ Recessive	Frequency Among Human Births		
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Hypercholesterolemia	Excessive cholesterol levels in blood lead to heart disease	Abnormal form of cholesterol cell surface receptor	Dominant	1/500		





Entire Genetic Code of a Bacteria



<u>PGD</u>

Pre-

Implantation

Genetic

Diagnosis

DNA Fingerprinting

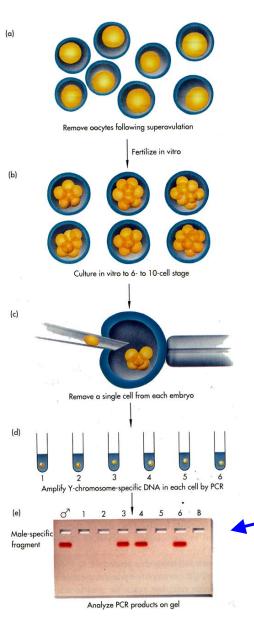


Cloning: Ethical Issues and Future Consequences



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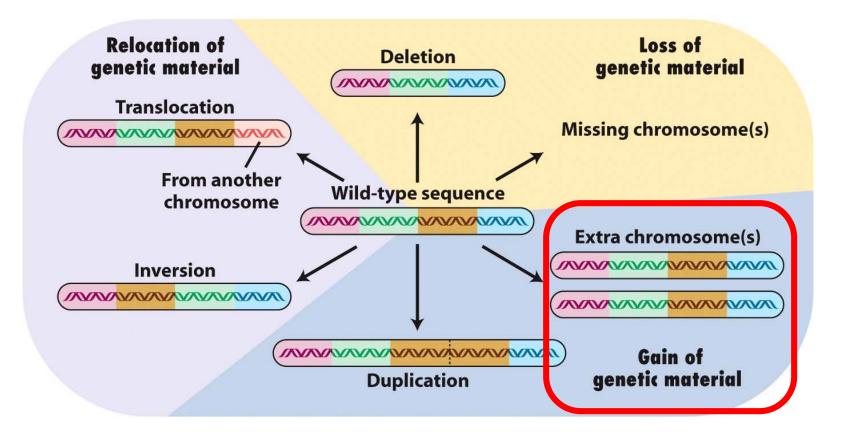
PCR Can Be Used To Analyze Mutant Gene in A Single Embryo Cell <u>Before</u> Pregnancy



What is The Implication of This Procedure Considering That The Human Genome Has Been Sequenced?

Sex Determination in 8-cell Embryo!

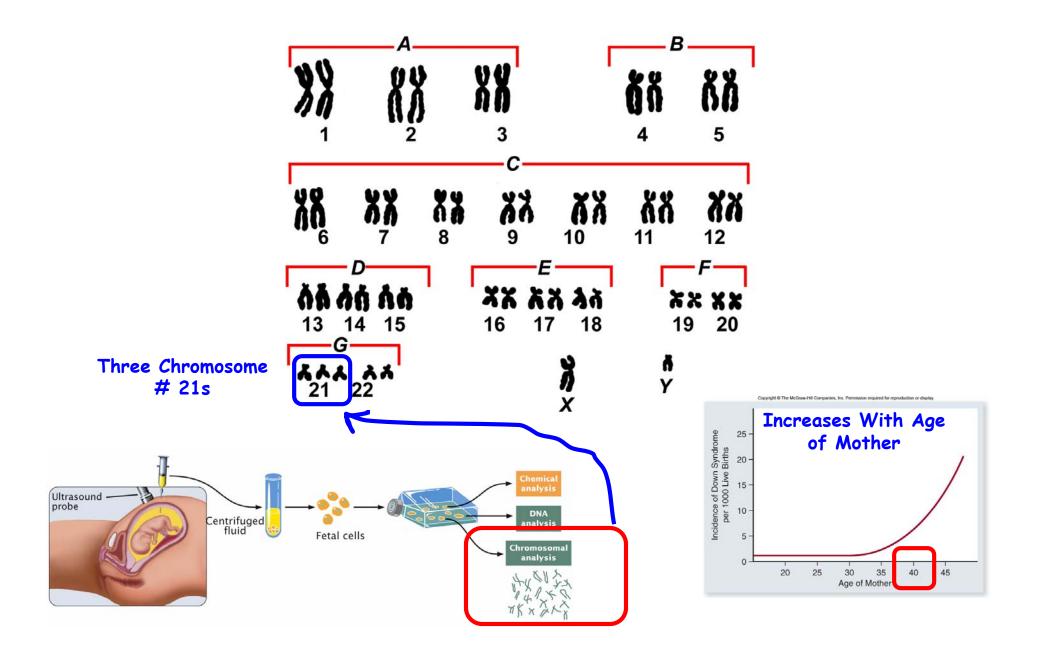
"Mutations" Can Also Occur By Large Chromosomal Changes



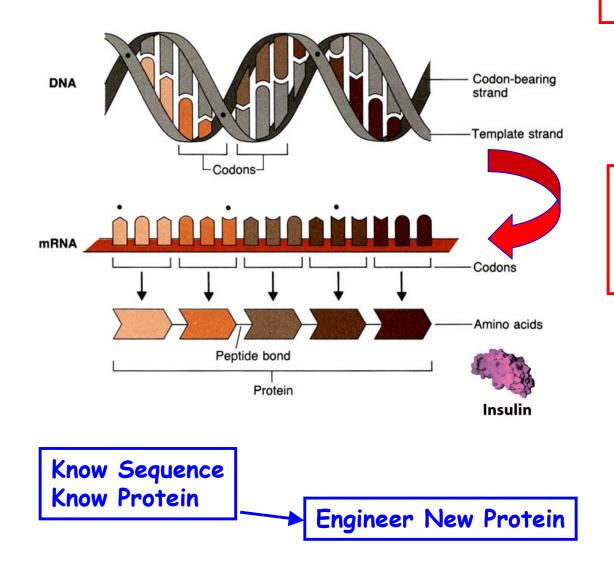
These changes affect many genes!

e.g. Down's Syndrome (3 Chromosome #21s)

A Down's Syndrome Karyotype



⁽²⁾ How Does A Gene Lead To A Phenotype?

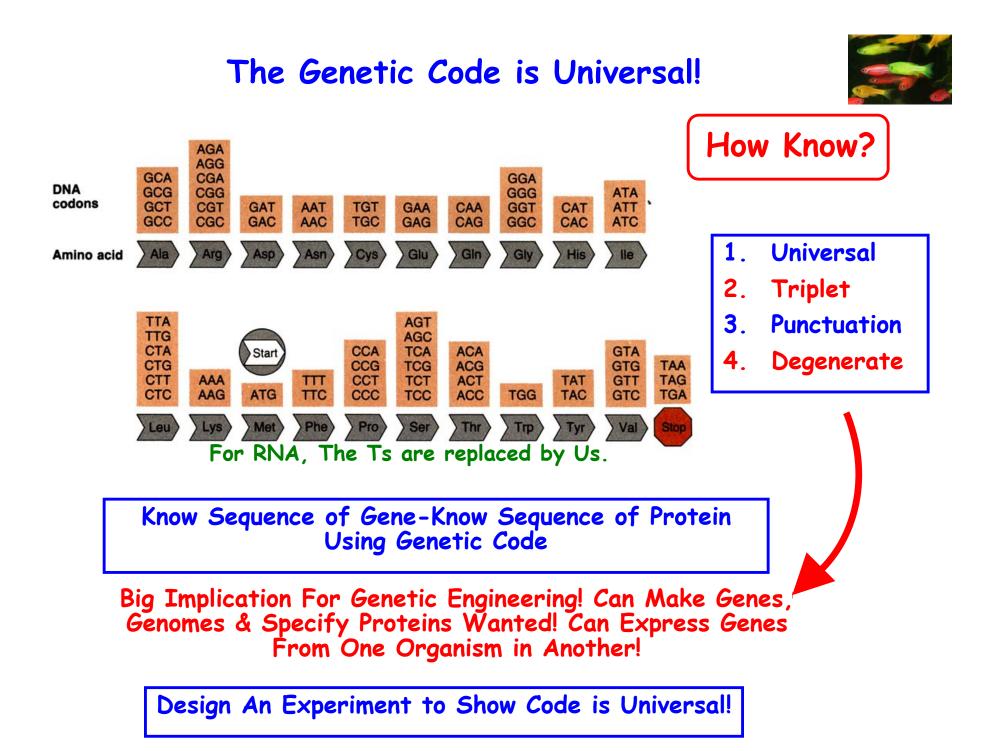


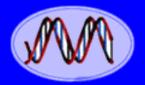
① mRNA Synthesized by Transcription

- Complementary to Transcribed, Non-Sense Strand
- Same Sequence As Sense Strand
- ② mRNA Translated into Protein by Translation of The Genetic Code

Genetic Code on mRNA Translated to Protein Sequence

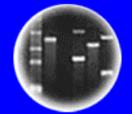
∴ Sequence of Gene Sequence of mRNA Sequence of Protein







Entire Genetic Code of a Bacteria



DNA Fingerprinting

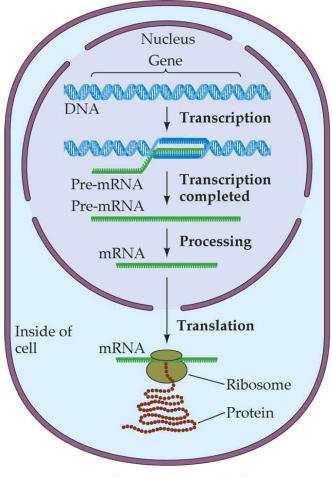


Cloning: Ethical Issues and Future Consequences



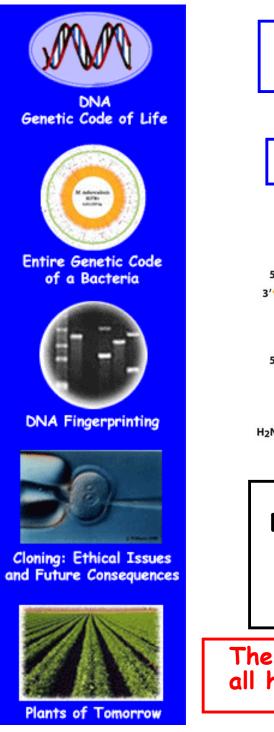
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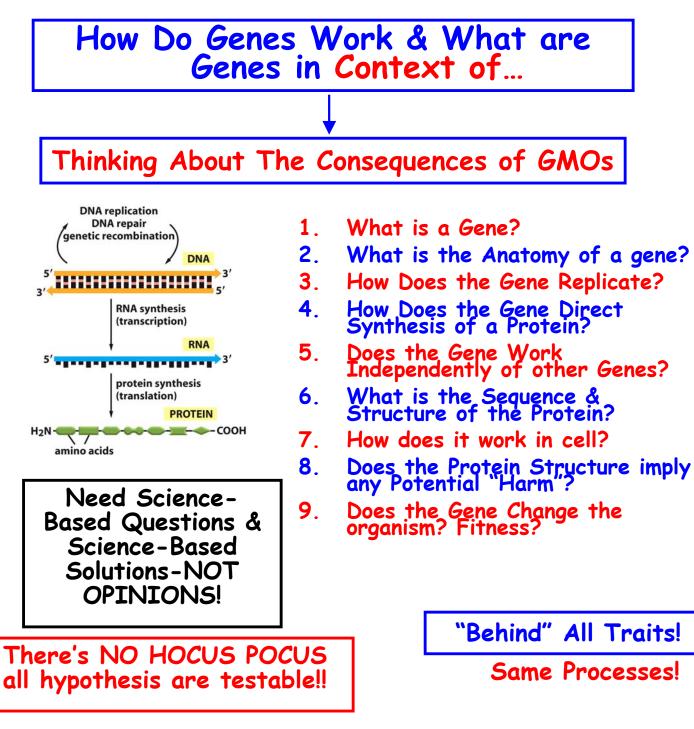
An Elaborate Cellular Machinery Requiring Thousands Of Genes is Required To Produce Proteins Encoded By Specific Genes!!

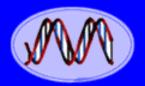


It takes Genes to Express (and Replicate) A GENE!!!

LIFE: THE SCIENCE OF BIOLOGY, Seventh Edition, Figure 14.1 Eukaryotic mRNA Is Transcribed in the Nucleus but Translated in the Cytoplasm © 2004 Sinauer Associates, Inc. and W. H. Freeman & Co.

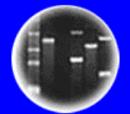








Entire Genetic Code of a Bacteria



DNA Fingerprinting



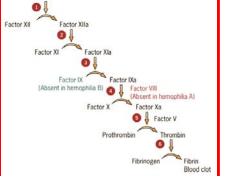
Cloning: Ethical Issues and Future Consequences



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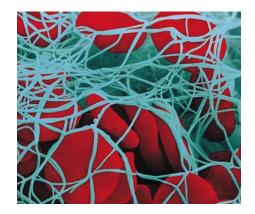
The Nuts & Bolts of Genetic Engineering

The Factor VIII Story: From Gene To Genetic Screening To Drug

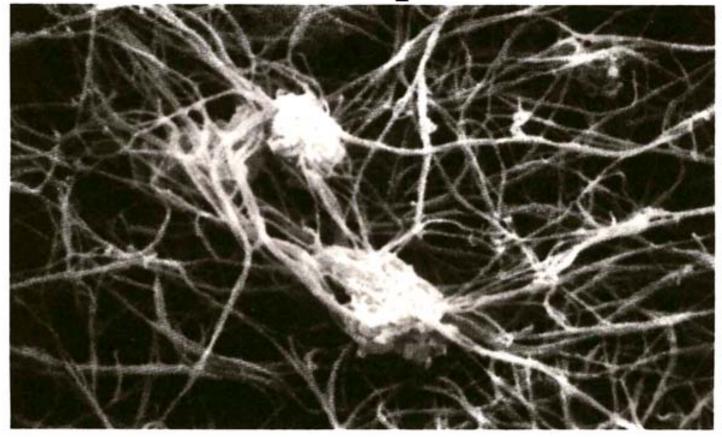


Wound





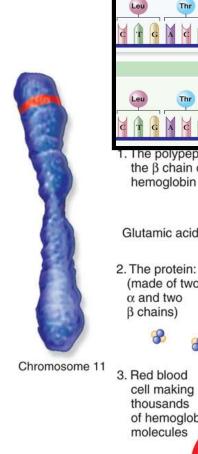
The Molecular Genetics of Hemophilia (Potentially Lethal Disease)

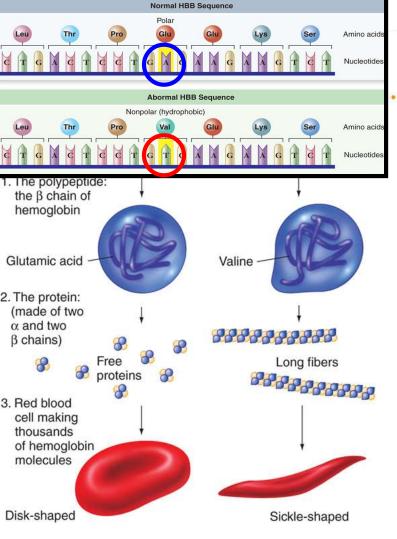


FIBRIN STRANDS stabilize a blood clot at the site of a wound by trapping the platelets that form the bulk of the clot. The electron micrograph, which was made by Jon C. Lewis of Wake Forest University, shows a clot formed in a suspension of platelets and fibrin. A clot in the bloodstream is the result of a complex cascade of enzymatic reactions culminating in the conversion of fibrinogen, a soluble protein, into insoluble fibrin strands. In hemophiliacs a crucial protein in the blood-clotting cascade is either missing or defective.

A Case Study of Cloning Genes and mRNAs Reference: Lawn & Vehar, Sci. Amer., January, 1986

Human Genetic Disorders Occur As A Result of Mutations: Change Code-Alter Protein





(b) Sickle-cell anemia is pleiotrophic

Si	ckling of red blood cell \downarrow	s —
Rapid destruction of sickle cells	Clumping of cells; interference with circulation	Accumulation of red blood cells in spleen
Anemia	tocal failures in blood supply ↓	Finlargement and damage to spleen
Fatigue, heart damage, overactivity of bone marrow	Damage to heart, kidney, muscle/joints, brain, lung, gastrointestinal tract	

(c) β-chain substitutions/variants

	Amino-acid position									
	1	2	3	· 6	7	· 26 ·	·· 63 ·	67.	·125·	146
Normal (HbA)	Val	His	Leu	Glu	Glu	Glu	His	Val	Glu	His
HbS	Val	His	Leu	Val	Glu	Glu	His	Val	Glu	His
HbC	Val	His	Leu	Lys	Glu	Glu	His	Val	Glu	His
HbG San Jose	Val	His	Leu	Glu	Gly	Glu	His	Val	Glu	His
HbE	Val	His	Leu	Glu	Glu	Lys	His	Val	Glu	His
HbM Saskatoon	Val	His	Leu	Glu	Glu	Glu	Tyr	Val	Glu	His
Hb Zurich	Val	His	Leu	Glu	Glu	Glu	Arg	Val	Glu	His
HbM Milwaukee 1	Val	His	Leu	Glu	Glu	Glu	His	Glu	Glu	His
HbDβ Punjab	Val	His	Leu	Glu	Glu	Glu	His	Val	GIn	His

Sickle-Cell Anemia

Hemophilia Has Been Known As An Inherited Disease For >2500 Years!

Old Testament-Circumcisions Royal Family-Europe





First Reference to Hemophilia is in the Old Testament

Genesis 17:10-14

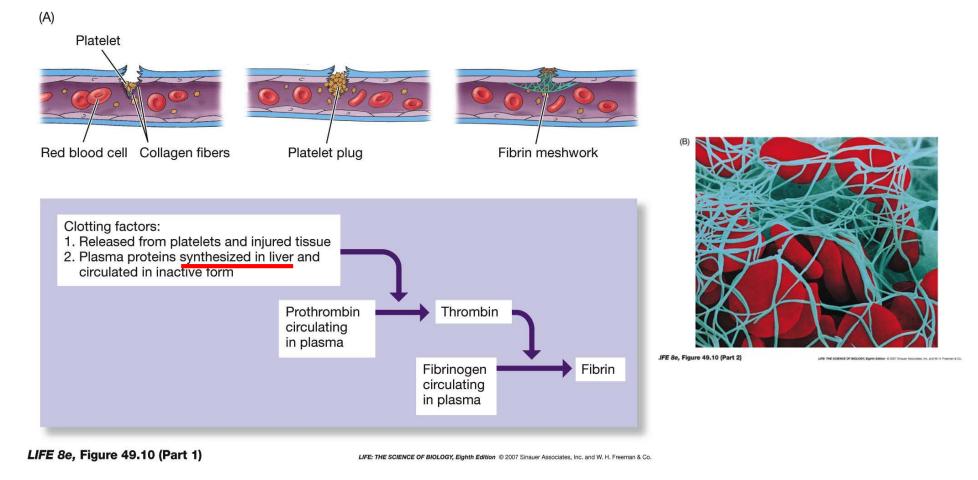
"This is My covenant that you shall keep between Me and you and your descendants after you: every male among you shall be circumcised. You shall circumcise the flesh of the foreskin......At the age of eight days every male among you shall be circumcised throughout your generations......an uncircumcised male...that soul shall be cut off from its people, he has invalidated My covenant.'



The Talmud also makes reference to families in whom children have died as a result of circumcision (Babylonian Talmud, Chapter Yevamoth p64b) [6]. Should a mother lose two children or should two sisters lose a child each after circumcision, subsequent children of the woman, the two sisters or of any other sisters of the same family should not be circumcised until they are older, or possibly not at all. This is thought to be the earliest reference to haemophilia; it was recognized in the Talmud that this condition was transmitted by the mother.

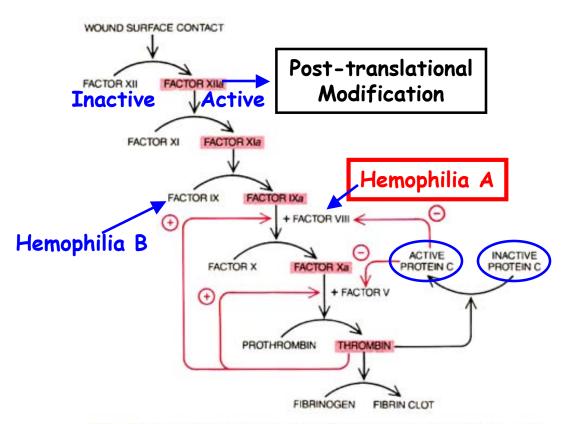
Abraham was circumcised at 93 and gave birth to Isaac at 99. His wife - Sarah - was 90!

A Cascade Of Events After Wounding Leads to A Fibrin Clot



Clotting Factors Such As Factor VIII Play A Critical Role in This Process

How Does Blood Clot After Wounding?



Eight Proteins/Genes Required:

- 1. Factor VII
- 2. Factor XI
- 3. Factor IX
- 4. Factor VIII
- 5. Factor X
- 6. Protein C
- 7. Prothrombin
- 8. Fibrinogen

CLOTTING CASCADE begins when cell damage at a wound somehow activates the enzyme factor XII; it ends with the conversion of fibrinogen into fibrin by thrombin. At each step an inactive protein is converted into a protease, or protein-cutting enzyme (color), which activates the next protein. Some steps require cofactors such as factors VIII and V. The cascade includes positive- and negative-feedback loops (colored arrows). Thrombin activates factors VIII and V; it also deactivates them (by activating protein C), which helps to halt clotting. Some 85 percent of hemophiliacs lack factor VIII. The rest lack factor IX.

ATryn[®] 2009

Anti-Thrombin??

Cascade

→Anti-Thrombin Deficiency (At-III) genetic disease

What Happens If Any Of These Proteins Or Genes Are Mutated?



J

Hemophiliacs Have Mutations In Either Factor VIII or Factor IX Genes

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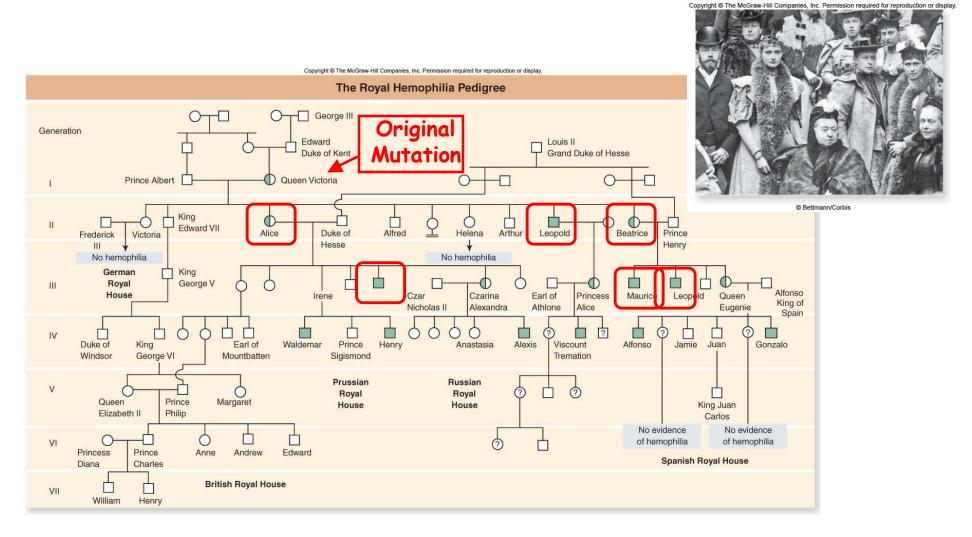
TABLE 13.2 Some Important Genetic Disorders					
Disorder	Symptom	Defect	Dominant/ Recessive	Frequency Among Human Births	
Cystic fibrosis	Mucus clogs lungs, liver, and pancreas	Failure of chloride ion transport mechanism	Recessive	1/2500 (Caucasians)	
Sickle cell anemia	Blood circulation is poor	Abnormal hemoglobin molecules	Recessive	1/600 (African Americans)	
Tay–Sachs disease	Central nervous system deteriorates in infancy	Defective enzyme (hexosaminidase A)	Recessive	1/3500 (Ashkenazi Jews)	
Phenylketonuria	Brain fails to develop in infancy	Defective enzyme (phenylalanine hydroxylase)	Recessive	1/12,000	
Hemophilia	Blood fails to clot	Defective blood-clotting factor VIII	X-linked recessive	1/10,000 (Caucasian males)	
Huntington disease	Brain tissue gradually deteriorates in middle age	Production of an inhibitor of brain cell metabolism	Dominant	1/24,000	
Muscular dystrophy (Duchenne)	Muscles waste away	Degradation of myelin coating of nerves stimulating muscles	X-linked recessive	1/3700 (males)	
Hypercholesterolemia	Excessive cholesterol levels in blood lead to heart disease	Abnormal form of cholesterol cell surface receptor	Dominant	1/500	

Hemophilia A	Defective Factor VIII Gene	1/10,000 males
Hemophilia B	Defective Factor IX Gene	1/30,000 males

Hypothesis For High Frequency in Males?

Both Genes On X-Chromosome $9 \rightarrow 3'$'s

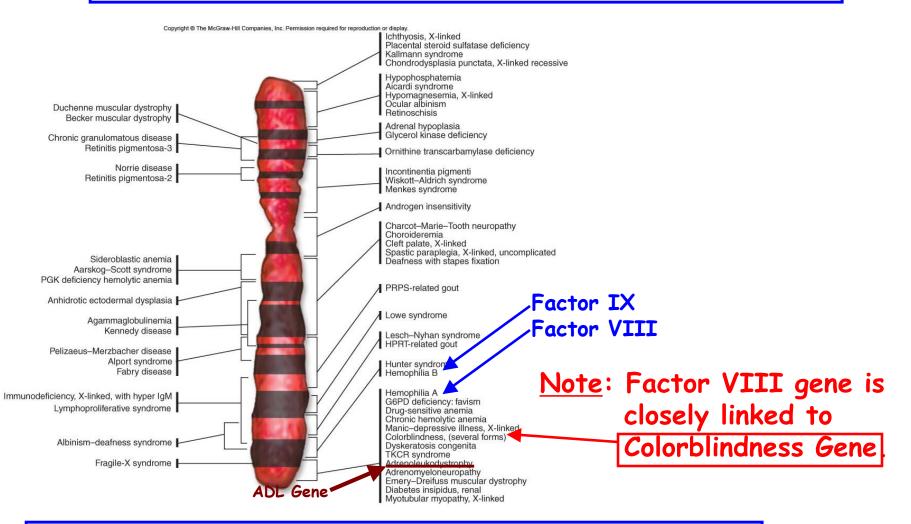
Hemophilia A and B Genes (Traits) Are Sex Linked



Note: 1. Males Obtain Detective Gene From Mothers

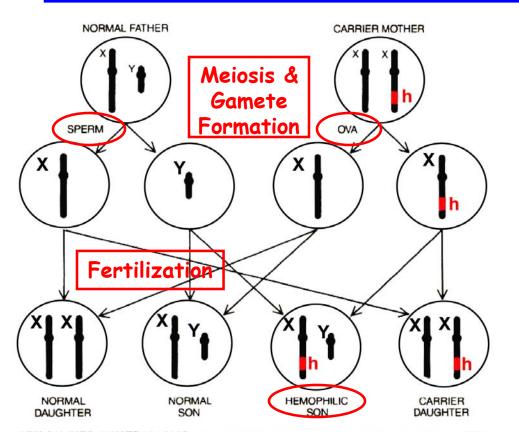
2. 50% of Sons Of A Maternal Carrier Have The Defective Gene

Factor VIII and Factor IX Genes are Closely Linked on the X Chromosome



The X chromosome has ~1500 Genes (2008) and 150,000,000 bp (150 Mb)

Hemophilia A and B Inheritance



Meiosis Gametes n Fertilization

Children

Parents

2n

> 2n

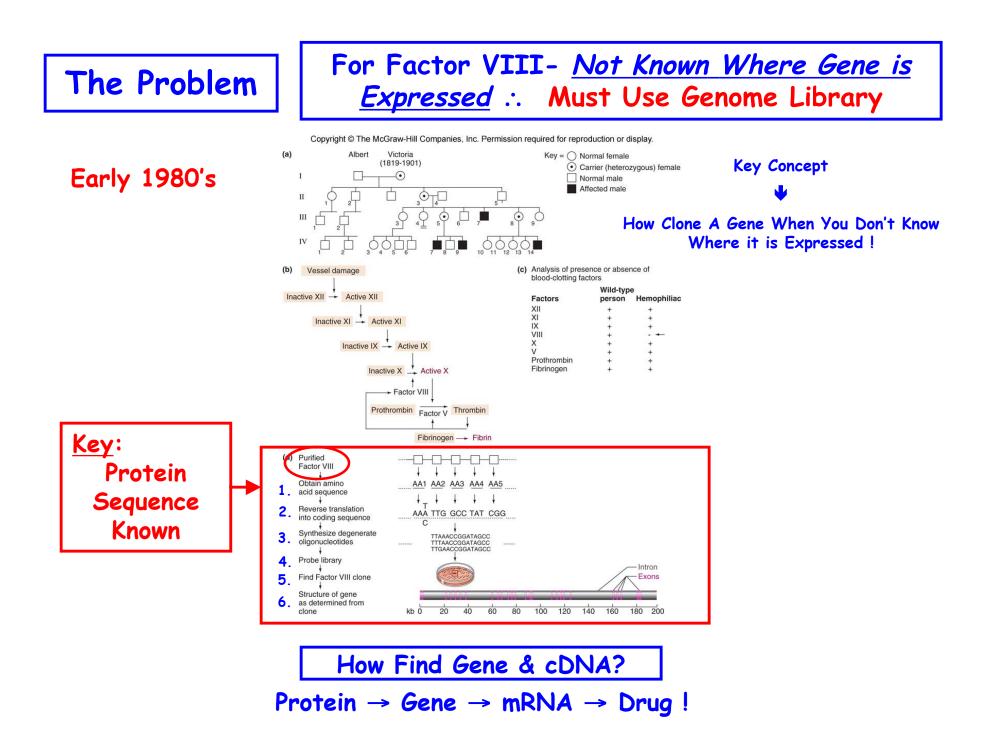
SEX-LINKED INHERITANCE of hemophilia results from the location of the factor VIII gene on the X chromosome. A male carrying a mutant factor VIII gene lacks normal factor VIII and is hemophilic. A female carrier is protected by the normal gene on her second X chromosome, but half of her daughters will be carriers and half of her sons will be hemophilic. In the case of a hemophilic father (not shown), his sons will not be hemophilic, because they receive his Y (not his X) chromosome, but his daughters will be carriers.

Sex-Linked Inheritance ♀ Carriers → 1/2 Sons + No Daughters! Only One X-Chromosome is ♂

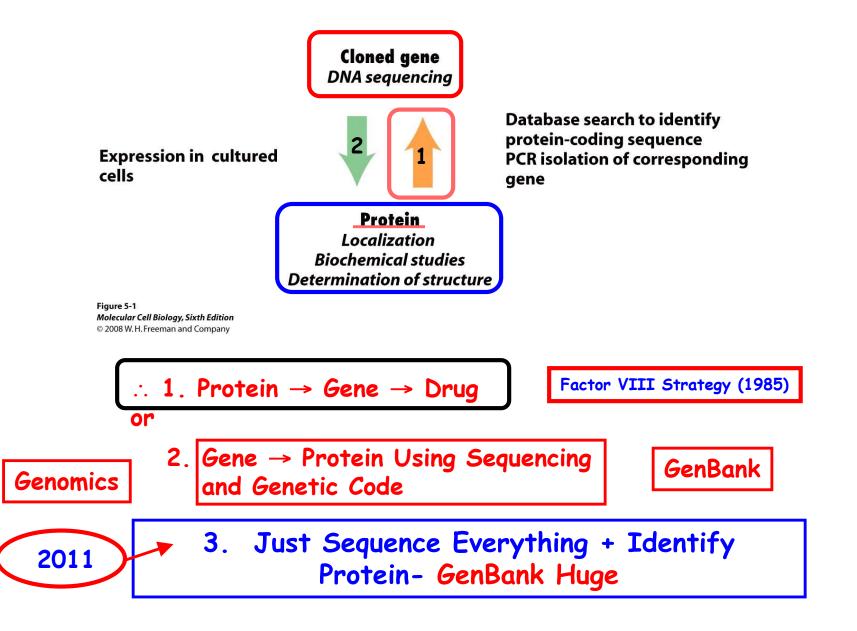
What Was Known About Factor VIII Before Gene Cloned?

- 1. Blood Protein (But Perhaps Synthesized Elsewhere!)
- 2. Could be purified in small amounts from >20 Liters of human blood +cow blood + pig blood
- 3. Short Stretch of <u>Proteins</u> Sequenced = Known Protein Sequence!
- ◆4. Hemophilia A could be treated by <u>blood transfusions</u> from normal individuals, ∴ clotting factor <u>in blood</u>.

:. How to go From Protein to Gene



Knowledge of the Protein Sequence and the Genetic Code Makes it Possible to Identify a Gene

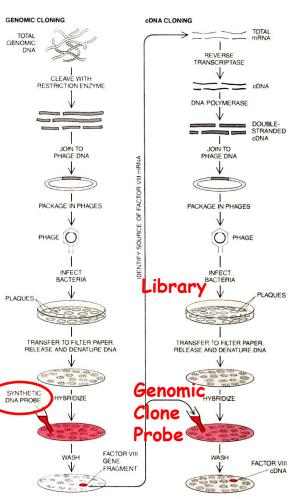


Steps Required to Clone Factor VIII Gene and cDNA

1. Make Genome Library Because Factor VIII Gene in Genome!

Gene

- 2. Purify Protein from Blood- that's where it works (wasn't known where made)
- 3. Reverse Translate using the genetic code a portion of the protein sequence
- 4. Synthesize a DNA probe complementary to Factor VIII gene corresponding to protein sequence
- 5. Screen Genome Library Entire Gene on The Clone?



cDNA

- 1. Use Gene probe to screen cDNA library for Factor VIII cDNA clone
- 2. How know what mRNA to use to make cDNA library?
- 3. Use gene probe to probe RNA blots containing mRNA from all major organs (liver, kidney, blood, etc.)
- 4. Find Factor VIII mRNA in livermale, liver- secrete into blood
 - Why Need cDNA? Story continued

Want cDNA to Manufacture Factor VIII as a Drug to Treat Hemophilia A!

Step One

How to Construct a Human Genome Library to Find the Factor VIII Gene?

If It is Not Known Where Gene is Active Can "Look" to Genome Instead of mRNA to Find + Clone Gene!

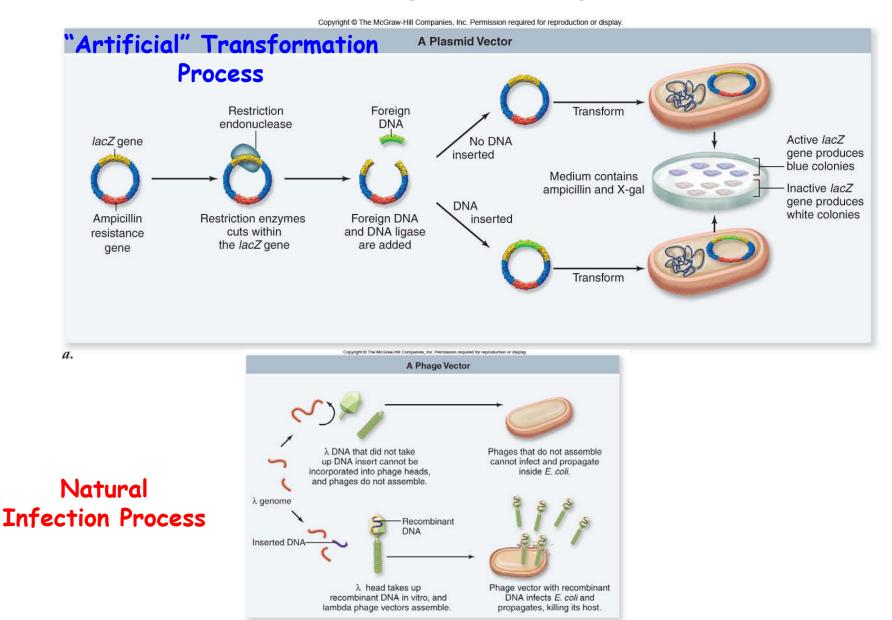
Vectors Used in Genetic Engineering Have Similar Conceptual Properties But are Used in Different Situations

Vector Type	Maximum Insert Size (kb)	Applications	Limitations
Bacterial plasmid vectors (circular)	~6-12	DNA cloning, protein expression, subcloning, direct sequencing of insert	Restricted insert size; limited expression of proteins; copy number problems; replication restricted to bacteria
	DNA	DNA	restricted to bacteria
Bacteriophage vectors (linear)	~25	cDNA, genomic and expression libraries	Packaging limits DNA insert size; host replication problems
Cosmid (circular)	~35	cDNA and genomic libraries, cloning large DNA fragments	Phage packaging restrictions; not ideal for protein expres- sion; cannot be replicated in mammalian cells
Bacterial artificial chromosome (BAC, circular)	~300	Genomic libraries, cloning large DNA fragments	Replication restricted to bacteria; cannot be used for protein expression
Yeast artificial chromosome (YAC, circular)	200-2,000	Genomic libraries, cloning large DNA fragments	Must be grown in yeast; cannot be used in bacteria
Ti vector (circular)	Varies depending on type of Ti vector used	Gene transfer in plants	Limited to use in plant cells only; number of restriction sites randomly distributed; large size of vector not easily manipulated

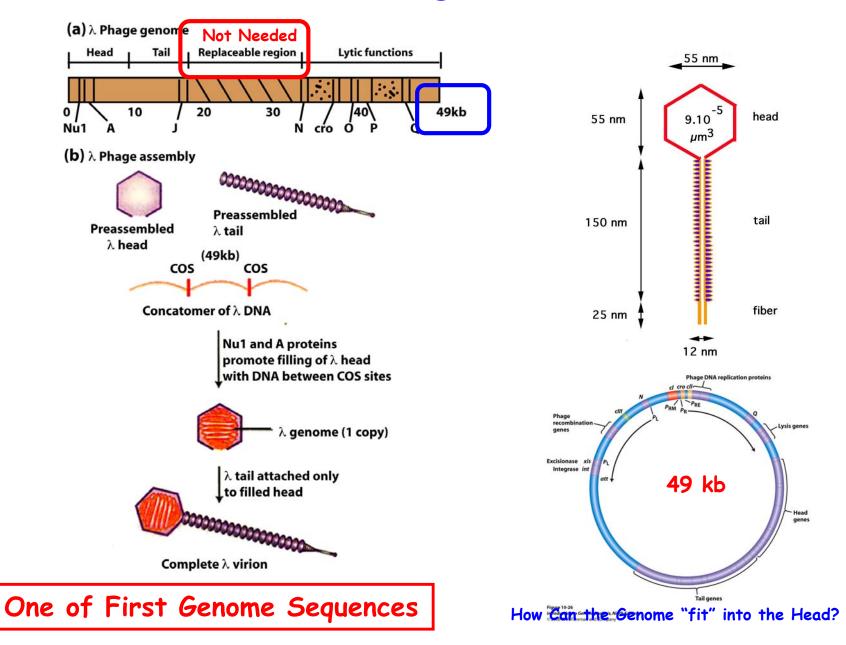
Plasmids vs. Bacteriophage Vectors

- 1. Replicate
- 2. Selectable
- 3. Can be used to insert foreign genes/restriction sites
- 4. Easily isolated + transferred back to cells

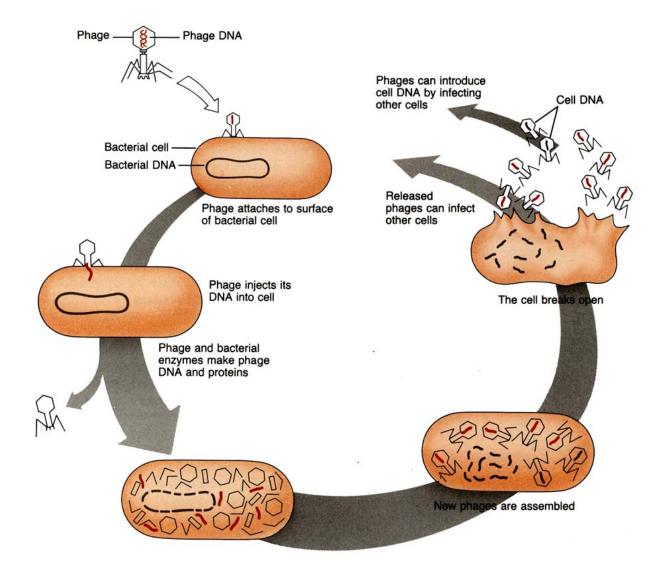
Plasmid vs. Bacteriophage Vectors for Cloning DNA Fragments



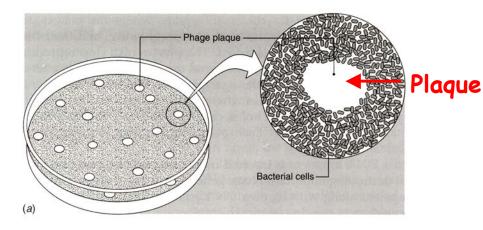
Structure of the λ Phage and Its Genome

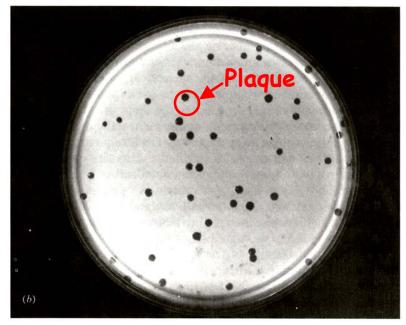


λ Phage Infects E.coli & Destroys (Lyses) cells



Lysed Cells Can Be Seen as Clear Plaques on Agar Plates





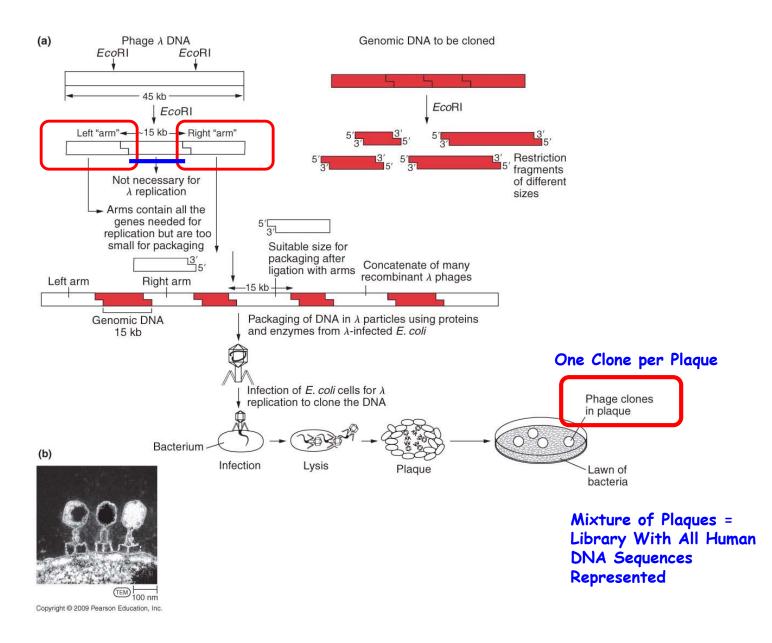
1. Each <u>Plaque</u> is a Virus Clone Representing One Viral Infection!

2. Selectable <u>Marker</u> is Bacterial Cell Destruction & Plaque Formation

Advantages of λ Virus as a Vector for Cloning DNA

- 1. Long DNA Segments can be Cloned (~20kb) Need fewer clones for whole Genome!
- 2. Can clone DNA Segments in Viral Genome & Self-Assemble with viral proteins into virus in a test tube!
 - ... Make Recombinant Viruses in the Lab!
- 3. Use "Natural" Infection process to Generate Large Number of Clones for a Eukaryotic Genome Library.
 Much higher efficiency for getting recombinant DNA → bacterial cells compared with DNA transformation.
 - ... set more clones per amount of recombinant DNA!

Using a Bacterial Virus To Clone the Human Genome



Cloning the Human Genome and Screening for the Factor VIII Gene

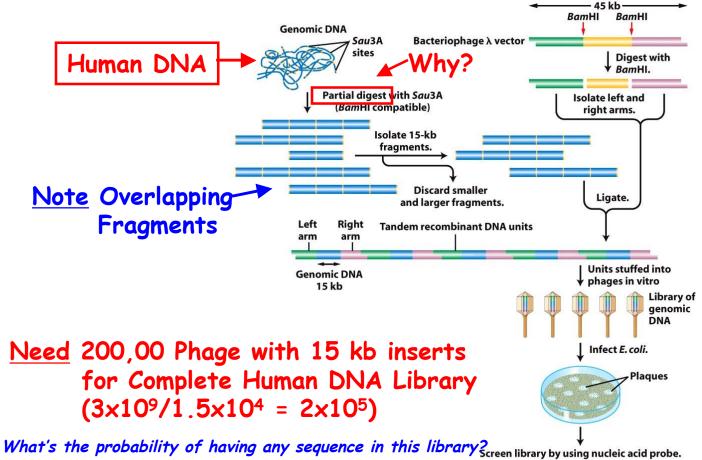


Figure 20-6 Introduction to Genetic Analysis, Ninth Edition © 2008 W.H. Freeman and Company

<u>Why</u> Partial Digestion? <u>An Important Concept</u>! <u>What</u> is Complete & Partial Digestion?

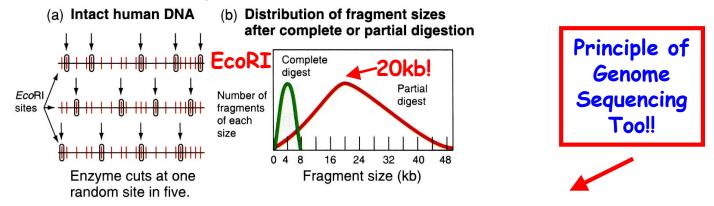
An EcoRI Restriction Enzyme Site is Found Only Once in the Human Genome:

- a. Yes
- b. No

What is the Purpose of Partial Digestion of Human DNA?

Sau 3A= 4bp= ⁵ 'GATC ^{3'}	∴ 1 site every 280bp if digest to completion = 1×10 ⁷ DNA fragments
Eco RI= 6bp= ⁵ GAATTC ³	∴ 1 site every 3100 bp if digest to completion (cleaves every site) = <u>972,000</u> DNA fragments

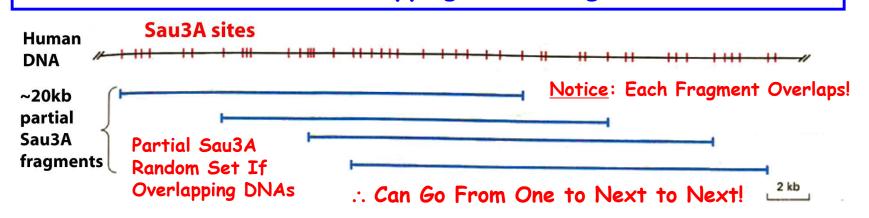
- 1. Complete Digestion Produces fragments that are too small to clone in λ virus (need 20Kb)
- 2. Complete Digestion would create huge genome libraries with large # clones to screen
- 3. Complete Digestion would break up genes of different DNA fragmentsparticularly if <u>human genes big</u>- ∴ would have one gene on many different clones- parts separated !
- 4. Complete Digestion provides no way to find <u>neighbors</u> of clones in genome- what's next to gene in chromosome!



Partial Digestion Produces A series of Large, Overlapping DNA Fragments/ Clones

Can connect one clone with another!! Build up clones of each chromosome!!

Constructing a Human Genome Library by Partial Digestion Creates a Set of Overlapping DNA Fragments/ Clones



... An overlapping set for each of the 24 chromosomes would allow clones to be ordered from beginning to end by restriction mapping because each chromosome contains one DNA molecule !

(B)

15

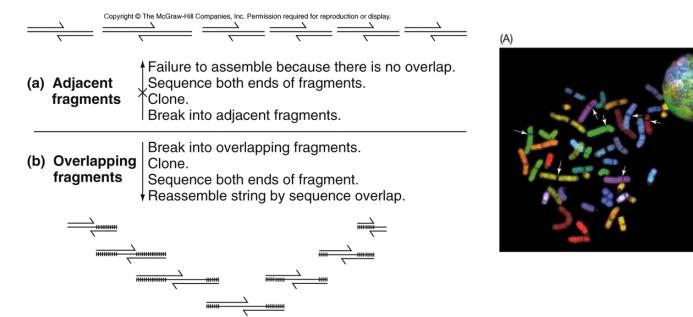
20

19

16

22

21



Step Two

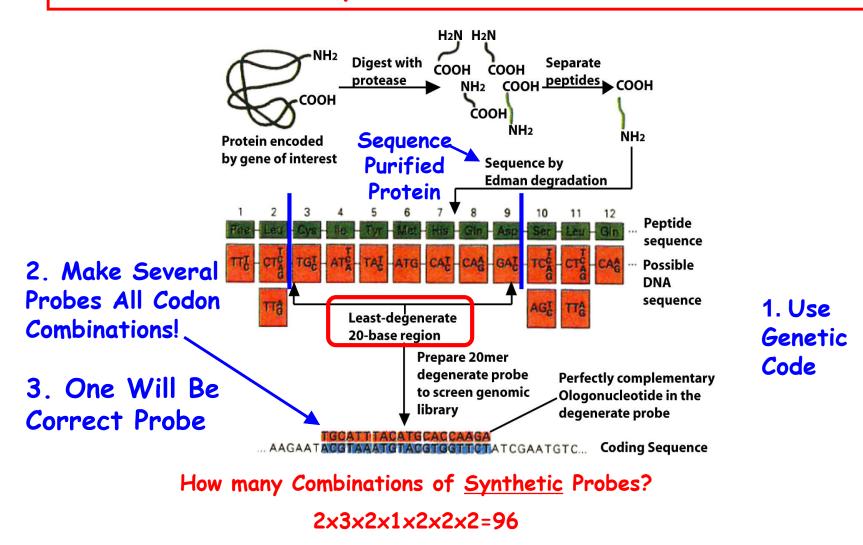
How Find the Factor VIII Gene in a Human Genome Library?

A Specific Gene Can Be Identified in a Genome Library if the Amino Acid Sequence of its Protein is Known Because of the :

- a. Double Helical Structure of DNA
- b. Antisense Strand DNA Sequence
- c. Genetic Code
- d. Mutant Gene Phenotype

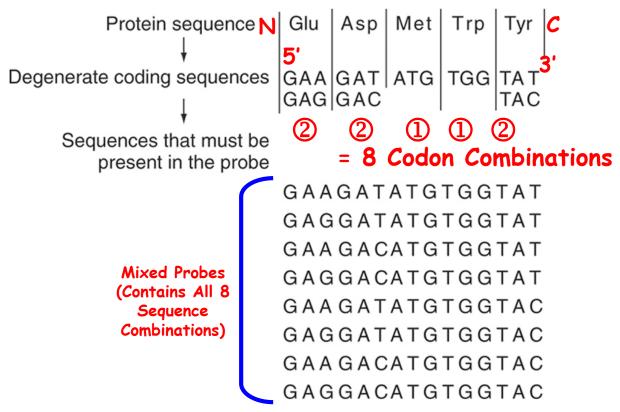
Factor VIII Protein → Gene

Using the Factor VIII Protein Sequence and Genetic Code as a Guide to Synthesize a Factor VIII Probe



Using the Genetic Code to go From Protein Sequence to Gene Sequence

Copyright © The McGraw-Hill Companies, Inc. Permission required for reproduction or display. (b) Synthesizing DNA probes based on reverse translation

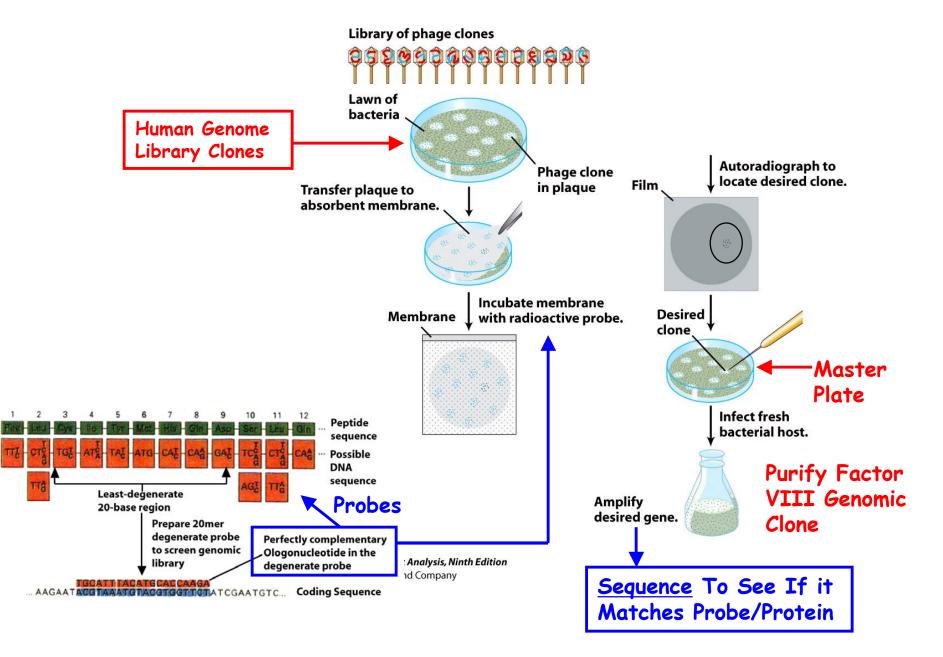


- 1. Need Amino Acid Sequence of Part of the Protein
- 2. Need DNA Sequences Representing all Codon Combinations
- 3. <u>Synthesize</u> DNA Sequence Probes!

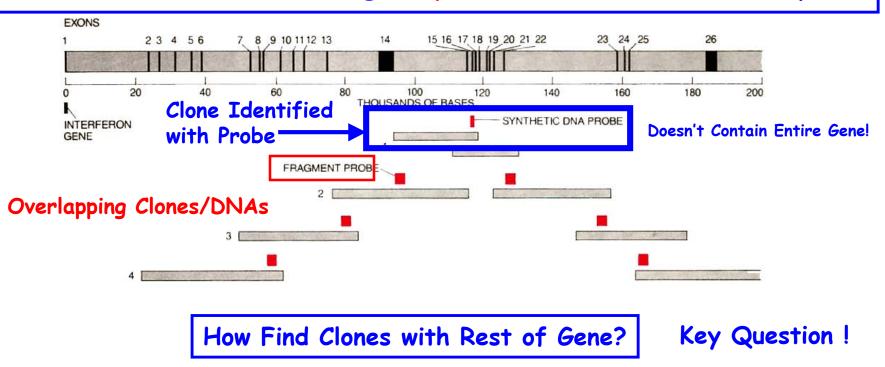
Probes Can Identify Genes in a Genome Library Because They Are: ?

- a. Synthetic
- b. Complementary to Specific DNA Sequences
- c. Contain the Correct Amino Acid Sequence
- d. Are Non-Radioactive

Finding The Factor VIII Gene Or Part of Gene!!



The Result-The Factor VIII Gene is Huge- 186,000 bp- The Probe Identified a Clone Containing Only One Part of Gene !!! <u>Why</u>?



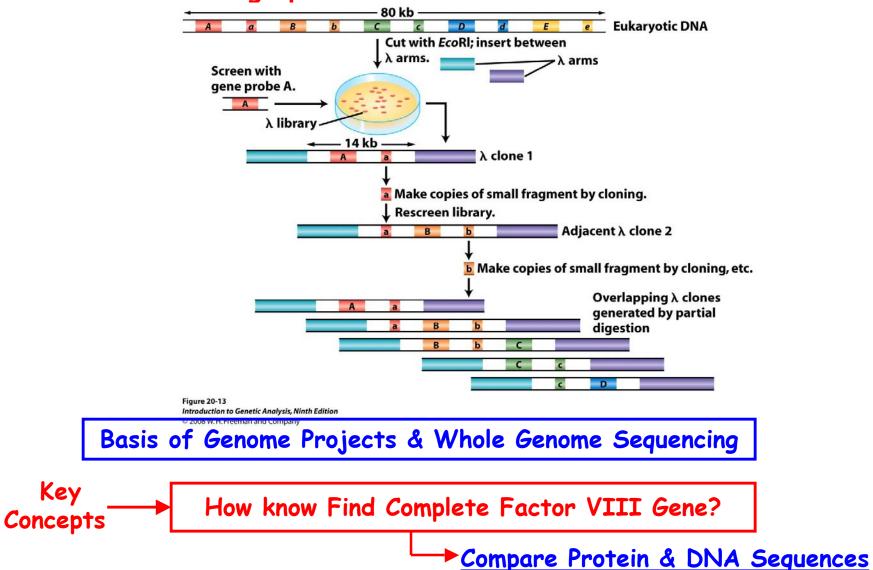
Remember - the library contains overlapping DNA clones .. can use one part of first clone to re-screen library & "walk" to other gene regions- using restriction maps & sequencing (compare with protein sequence) as guides!

Sequence -----> GenBank



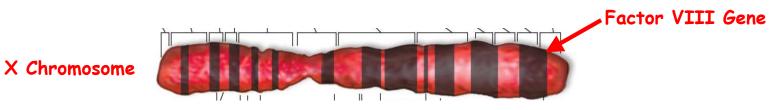
Finding the Entire Factor VIII Gene? Walking & Sequencing

Walking up and down Genes and Chromosomes

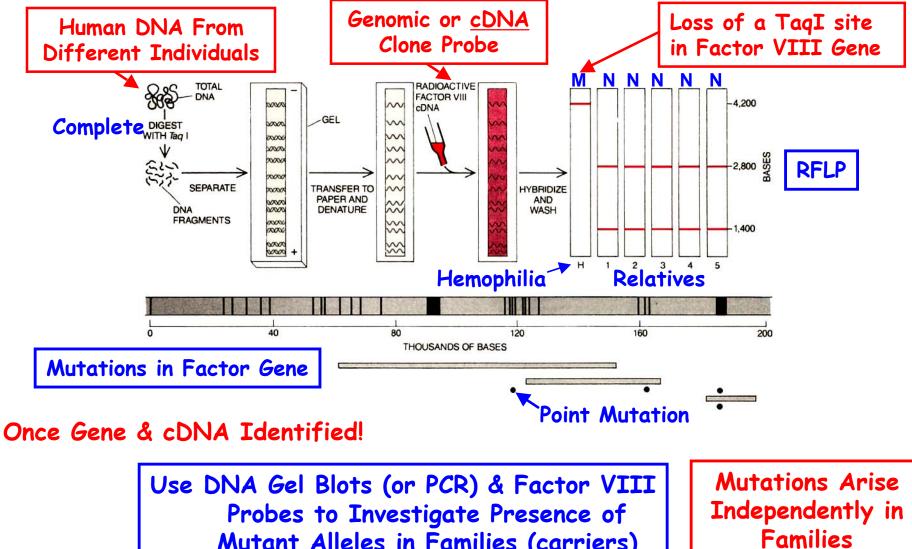


The Factor VIII Gene Was Found To Be Very Large

- 186,000 Nucleotides in Length (Won't Fit in One Phage Clone)
- · 25 Introns
- 9,000 Nucleotide Coding Sequence (cDNA)
- 2,351 Amino Acids in Protein



Factor VIII Gene Probes/ Sequence Can Be Used to Characterize Mutant Genes & Do DNA Testing for Carriers



Mutant Alleles in Families (carriers)

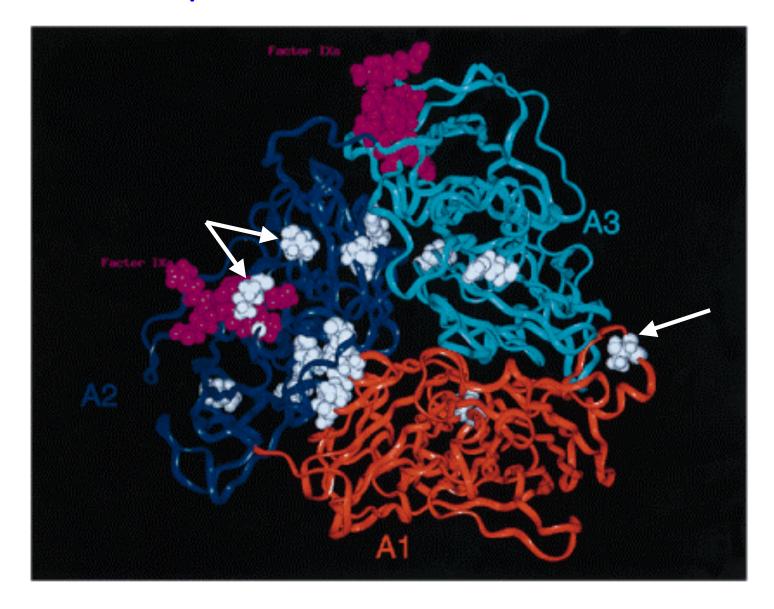
Factor VIII Mutations Occur Throughout the Gene

[Haemophilia 11, 481-491 (2005)]

VIII:C (%)	Family history	Consanguinity*	Inversion	Codon†	Mutation	Amino acid change	Exon	Conservation [‡]
L .	Sporadic	NC	Normal	51	$TTT \rightarrow TCTS$	Phe -> Ser	2	FFFF, identical
1.20	Sporadic	NC	Normal	80	$GTT \rightarrow GAT$	$Val \rightarrow Asp$	3	VVVV, identical
1	Sporadic	NC	Normal	102	$GGT \rightarrow GTT$	$Gly \rightarrow Val$	3	GGGG, identical
2	Sporadic	NC	Normal	104	$TCC \rightarrow CCCS$	Set -> Pro	3	SSSS, identical
6	Sporadic	NC	Normal	143	$GAG \rightarrow AAGS$	Glu → Lys	4	EEEE, identical
1	Sporadic	NC	Normal	233	delCA§	Thr \rightarrow fs (TGA-264)	6	
2.70	Inherited	NC	Normal	321	$GAA \rightarrow AAA$	$Ghu \rightarrow Lys$	8	EEEE, identical
1	Sporadic	NC	Normal	372	$CGC \rightarrow CAC$	Arg -> His	8	RRRR, identical
k	Inherited	NC	Normal	527	$CGG \rightarrow TGG$	Arg \rightarrow Trp	11	RRRR, identical
1	Sporadic	NC	Normal	52.8	$TGC \rightarrow TACS$	Cys -> Tyr	11	CCCC, identical
1	Inherited	NC	Normal	592	CAA -> TAA	Gln → Stop	12	QQQQ, identical
1	Inherited	NG	Normal	864	delGACA	Gly → fs [TAA-867]	14	**************************************
					insCAATTAAATGAGAA§			
t i	Sporadic	NC	Normal	948	insA§	Lys \rightarrow fs (TGA-984)	14	
1	Sporadic	NC	Intron 1	1107	AGG → TGGS	Arg -> Trp	14	RGKK, dissimilar
1	Sporadic	NC	Normal	1107	$AGG \rightarrow TGG$	$Arg \rightarrow Trp$	14	RGKK, dissimilar
t	Inherited	NC	Normal	1191-1194	delA	lle \rightarrow fs (TAG-1198)	14	
1.40	Sporadic	NC	Normal	1191-1194	insA	lle \rightarrow fs (TAA-1220)	14	
1	Sporadic	C	Normal	1227	delC§	Leu \rightarrow fs (TGA-1231)	14	
2.10	Sporadic	NC	Normal	1241	$GAC \rightarrow GAG$	$Asp \rightarrow Glu$	14	DGGE, similar
t -	Sporadic	NC	Normal	1392	1392dcl1418§	Pro \rightarrow fs (TAG-1446)	14	
1	Incrited	C	Normal	1.392	1392del14185	Pro \rightarrow fs (TAG-1446)	14	
1.5	Sporadic	NC	Normal	1441	insAS		14	
1	Incrited	C	Normal	1441	insA§			
1	Inherited	NC	Normal	1.502	CAG → TAG§	Gln → Stop	14	QREQ, dissimilar
1	Inherited	NC	Normal	1504	delGT§	Val \rightarrow fs (TGA-1517)	14	CINCRETE IN REAL MADE
1	Sporadic	NC	Normal	1535	$TGG \rightarrow TGA$	Trp → Stop	14	WLWM, dissimilar
hibitor 96 BU	San Sherrows					and an and the		
1	Sporadic	NC	Normal	1.571	$TAT \rightarrow TAAS$	Tyr \rightarrow Stop	14	Y-YY, dissimilar
1	Sporadic	NC	Normal	1.581	AAA -> TAAS	Lys -> Stop	14	KEKK, dissimilar
0.20	Sporadic	NC	Normal	1696	CGA → GGA	Arg -> Gly	14	RRRR, identical
.80	Sporadic	NC	Normal	1729	delA§	Gln \rightarrow fs (TAA-1752)	15	
1	Inherited	NC	Normal	1751	GAA → AAA§	$Ghu \rightarrow Lys$	15	EEEE, identical
1	Sporadic	NC	Normal	1775	$TTC \rightarrow TCC$	Phe \rightarrow Pro	16	FFFF, identical
t i	Sporadic	NC	Normal	1835	TGG → TGAS	Trp → Stop	16	WWWW, identical
7.60	Sporadic	C	Normal	1882	$ATC \rightarrow ATAS$	$IIe \rightarrow IIe$	17	IIII, identical
3	Inherited	C	Normal	1966	$OGA \rightarrow CAA$	Arg → Glu	18	RRRR , identical
1	Sporadic	NC	Normal	1966	$CGA \rightarrow TGA$	Arg -> Stop	18	RRRR, identical

Need To Screen Across the Gene for Markers -- Family Specific

Factor VIII Protein Structure & Positions Where Mutations Disrupt Protein Function and Lead to Hemophilia



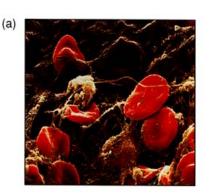
How is a Specific Gene Detected in Genome?

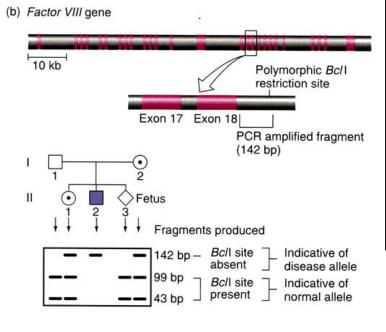
DNA can be Transferred "in situ" to paper & annealed with radioactive probes Solution passes through **RNA or DNA** gel and membrane to paper towels. Migration Paper towels Sponge 32P-labeled Electrophoresis size markers **DNA Blots!** Gel Salt Membrane solution Filter Gel DNA Hybridize with unique transferred nucleic acid probe. to membrane Probe Represents a Filter in "seal-a-meal" Cloned Fragment from Genome with a bag. Probe Remove hybridized to unbound Unique Sequence! complementary probe. sequence Expose X-ray film to membrane. == Autoradiogram

Figure 20-12 Introduction to Genetic Analysis, Ninth Edition © 2008 W. H. Freeman and Company

Using PCR and RFLPs (Markers) to Detect the Hemophilia A Disease Allele/ Gene

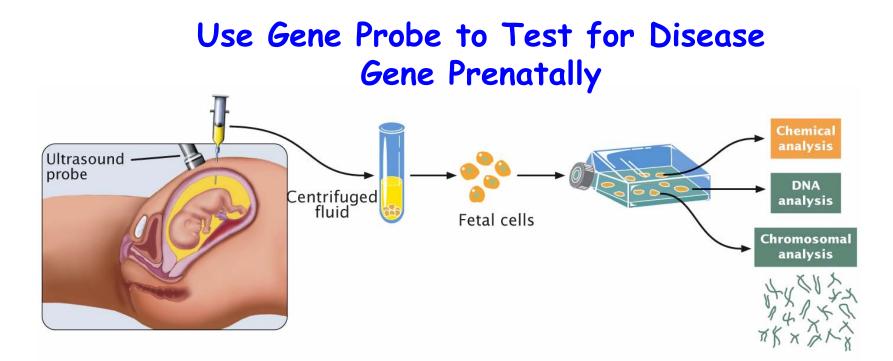
- 1. Use PCR to amplify a specific Factor VIII gene region
- Use restriction enzyme (BcL I) to distinguish between normal allele (1 site) & disease allele (no site)
 - = Normal allele
 - = Disease allele







Only Can Do This With a Knowledge of DNA Sequence of Wild-type (Normal) and Disease Genes (Can Vary family to Family)



Ultrasound Picture

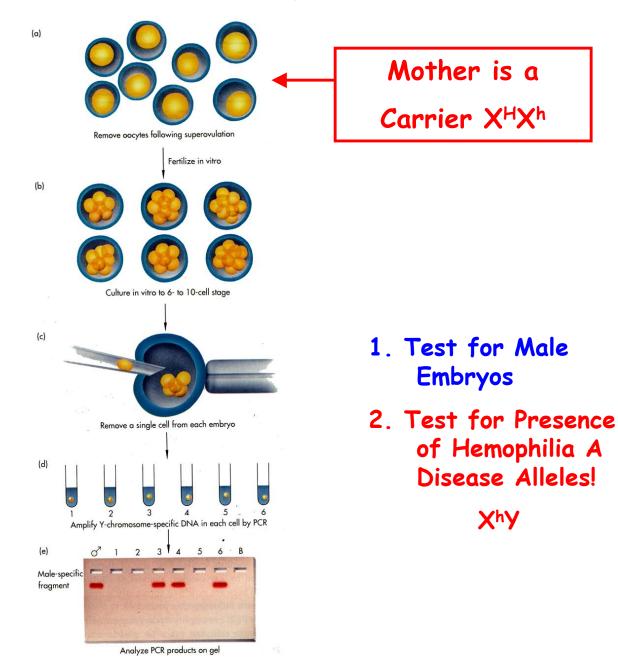


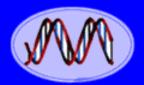
Fig_06-15 Genetics, Second Edition © 2005 W.H. Freeman and Company

Table 6.5	Examples of genetic diseases and disorders that can be detected prenatally and the techniques
	used in their detection

Disorder	Method of Detection
Chromosome abnormalities	Examination of a karyotype from cells obtained by amniocentesis or CVS
Cleft lip and palate	Ultrasound
Cystic fibrosis	DNA analysis of cells obtained by amniocentesis or CVS
Dwarfism	Ultrasound or X-ray; some forms can be detected by DNA analysis of cells obtained by amniocentesis or CVS
Hemophilia	Fetal blood sampling* or DNA analysis of cells obtained by amniocentesis or CVS
Lesch-Nyhan syndrome (deficiency of purine metabolism leading to spasms, seizures, and compulsory self-mutilation)	Biochemical tests on cells obtained by amniocentesis or CVS
Neural-tube defects	Initial screening with maternal blood test, followed by biochemical tests on amniotic fluid obtained by amniocentesis and ultrasound
Osteogenesis imperfecta (brittle bones)	Ultrasound or X-ray
Phenylketonuria	DNA analysis of cells obtained by amniocentesis or CVS
Sickle-cell anemia	Fetal blood sampling or DNA analysis of cells obtained by amniocentesis or CVS
Tay-Sachs disease	Biochemical tests on cells obtained by amniocentesis or CVS

Using PGD to Detect Hemophilia A Disease Alleles

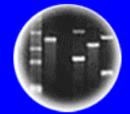




DNA Genetic Code of Life



Entire Genetic Code of a Bacteria



DNA Fingerprinting



Cloning: Ethical Issues and Future Consequences



Plants of Tomorrow

Genetic Screening Issues

•Why Screen For Genes? •When is a Test Accurate Enough? Mandatory or Voluntary Screening? •Who Should Be Tested? •Employer & Insurance Company Testing? Protection From Genotype Discrimination? •Testing for Genetic Diseases With No Cures?? •How Ensure Privacy & Confidentiality? •Obligations to Inform Others (Spouse/Sibling) of Genetic Disorder Knowledge?

•Genetic Databases??

Step Four

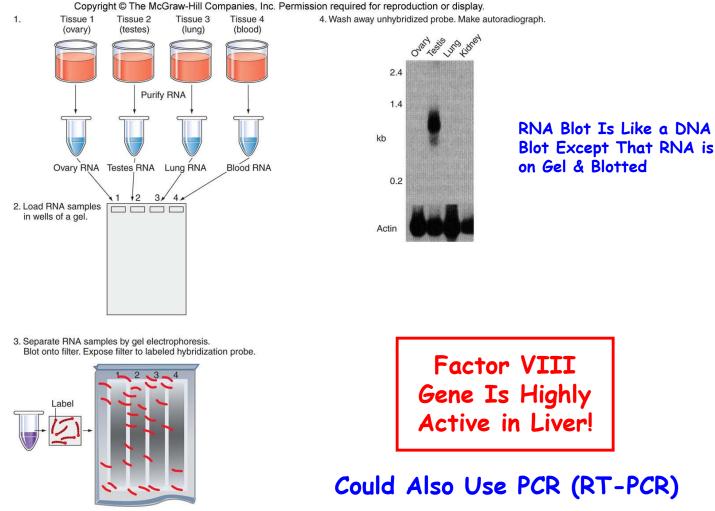
How Find Factor VIII mRNA to Generate a cDNA for Protein Production in Host Cells?

<u>Recall</u>: Eukaryotic Genes Provide Obstacles for Efficient Protein Production in Genetically Engineered Cells! <u>Reasons??</u>?

Making the Drug

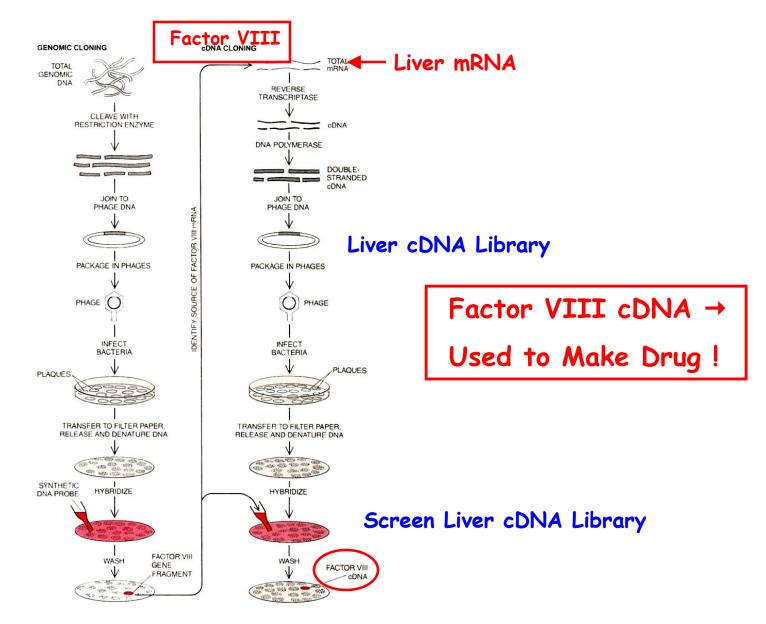
Need cDNA Not Gene

Factor VIII Gene Can Be Used to Find Out Where It is Active Using RNA Blots



(4): Reprinted with permission from Nature 1990 Jul 19; 346(6281):216-7, Sinclair et al. © 1990 Macmillian Magazines Limited

Using Factor VIII Gene Probe to Identify Factor VIII cDNA clone



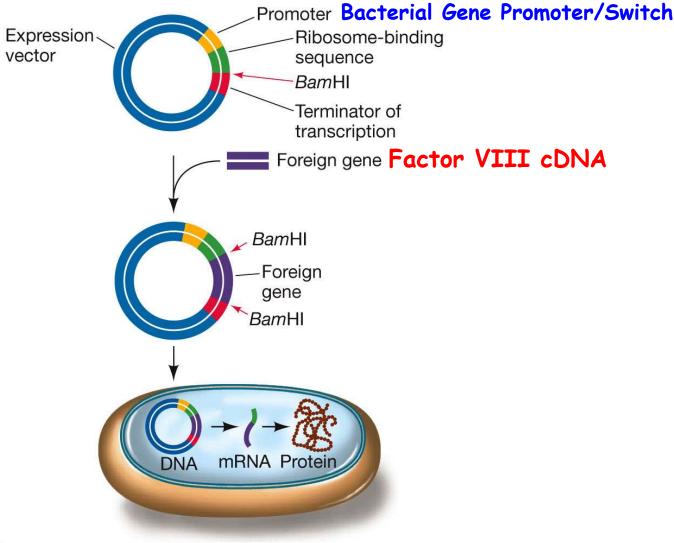
A cDNA is made by using:

- a. Primers
- b. mRNA
- c. DNA Polymerase
- d. All of above

The sequence of a cDNA clone is the same as:

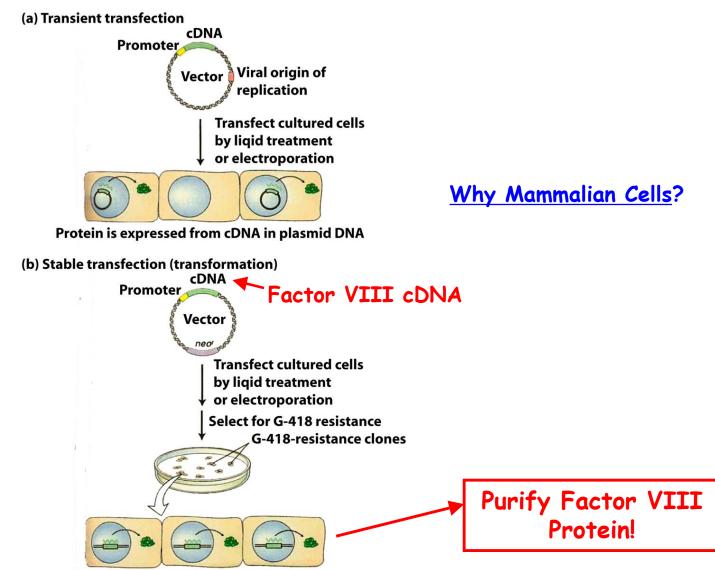
- a. The Sense Strand of the Corresponding Gene
- b. The mRNA Template
- c. The Antisense (Template Strand) of the Corresponding Gene
- d. The Sense and Antisense Strands of the Corresponding Gene Minus Introns

Use Expression Vector to Allow cDNA to Produce Protein in Host Cell



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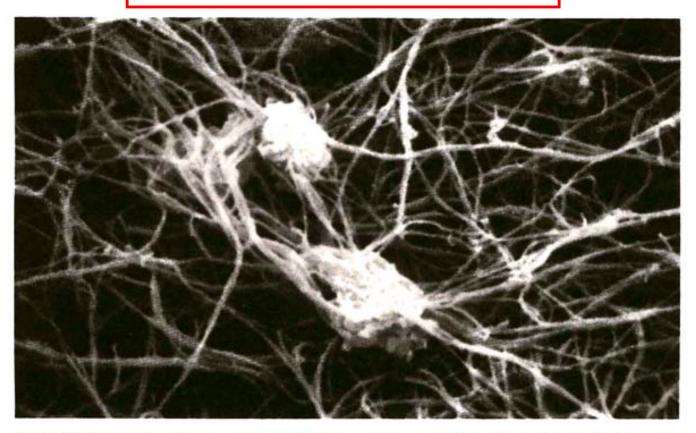
A Factor VIII Drug/"Cure" Making Factor VIII in Mammalian Cells



Protein is expressed from cDNA integrated into host chromosome

Using Factor VIII to Treat Hemophilia

Formation of a Blood Clot



FIBRIN STRANDS stabilize a blood clot at the site of a wound by trapping the platelets that form the bulk of the clot. The electron micrograph, which was made by Jon C. Lewis of Wake Forest University, shows a clot formed in a suspension of platelets and fibrin.

A clot in the bloodstream is the result of a complex cascade of enzymatic reactions culminating in the conversion of fibrinogen, a soluble protein, into insoluble fibrin strands. In hemophiliacs a crucial protein in the blood-clotting cascade is either missing or defective.

A Triumph of Genetic Engineering

Recombinant Factor VIII



Bayer Biological Products EU

(RAYER) Bayer HealthCare

Biological Products Division Search | Sitemap

Home

Recombinant Factor VIII

More Resources Haemophilia Centres in Europe

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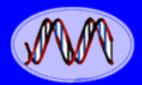
Recombinant factor VIII

Recombinant factor VIII (rFVIII) is the antihaemophilic factor A, obtained using recombinant DNA technology. With this technology, pure protein is synthesized in the laboratory instead of being extracted from blood. In the following pages, it will be explained in detail how the knowledge and analysis of DNA, using the new instruments of molecular genetics, have represented both the beginning



Factor VIII gene cloned in 1983

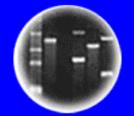
Factor VIII (recombinant) approved as drug in 1993! Ten years from gene → drug! (Off Patent in 2011)



DNA Genetic Code of Life



Entire Genetic Code of a Bacteria



DNA Fingerprinting



Cloning: Ethical Issues and Future Consequences



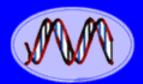
Plants of Tomorrow

A Patent on YOUR Factor VIII Gene!

United States Patent 5,618,788 April 8, 1997 Capon, et al. Preparation of functional human factor VIII and pharmaceutical treatment therewith Abstract Functional human factor VIII produced recombinantly is used in the treatment of human beings diagnosed to be deficient in factor VIII coagulant activity. Also provided are DNA solates and expression vehicles encoding functional human factor VIII, as well as transformed host cells and processes for producing human factor VIII by use of recombinant DNA echnology. Inventors: Capor; Daniel J. (San Mateo, CA), Lawn; Richard M. (San Francisco, CA), Vehar; Gordon A. (San Carlos, CA), Wood; William I. (San Mateo, CA) Assignee: Genentech, Inc. (South San Francisco, CA) Appl. No.: 07/570.096 Filed: August 20, 1990

An Individual Should Be Allowed to Patent the Factor VIII DNA Sequence:

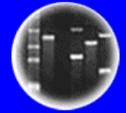
- a. Yes
- b. No



DNA Genetic Code of Life



Entire Genetic Code of a Bacteria



DNA Fingerprinting



Cloning: Ethical Issues and Future Consequences



Plants of Tomorrow

The Factor VIII Story -- A Summary

- 1. Purify Small Amounts of Factor VIII
- 2. Obtain Partial or Complete Amino Acid Sequence
- 3. Use the Genetic Code to Synthesize Degenerate DNA Probes
- 4. Isolate Factor VIII DNA Clones Complementary to Probe in Genome Library
- 5. Determine if Factor VIII Clones Contain the Complete Gene By Sequencing and Comparing With Protein Sequence
- 6. If Not, "Walk" to Obtain Overlapping DNA Clones That Collectively Contain the Factor VIII Gene
- 7. Sequence Clones To Determine Where the Factor VIII Gene Starts and Stops
- 8. Use Factor VIII Genome Probe to Find Out What Body Organ/Tissue Expresses the Factor VIII Gene
- 9. Make a cDNA Library From the Target Organ/Tissue and Isolate a Factor VIII cDNA Clone
- 10. Sequence the Factor VIII cDNA Clone and Compare With Factor VIII Gene Sequence to Map its Anatomy (I.e., introns, exons, swtiches) and Ensure That it Contains the Complete Protein Coding Sequence
- 11. Use Factor VIII cDNA and/or Genome Fragments as a Probe to Find RFLP Markers For Disease Alleles -- Or Sequence Disease Alleles to Find Relevant RFLP Markers By Comparison With Wild-Type Sequence
- 12. Insert Factor VIII cDNA Into an Expression Vector and Synthesize Factor VIII Protein in Host Cells (e.g., Mammalian Cells)